



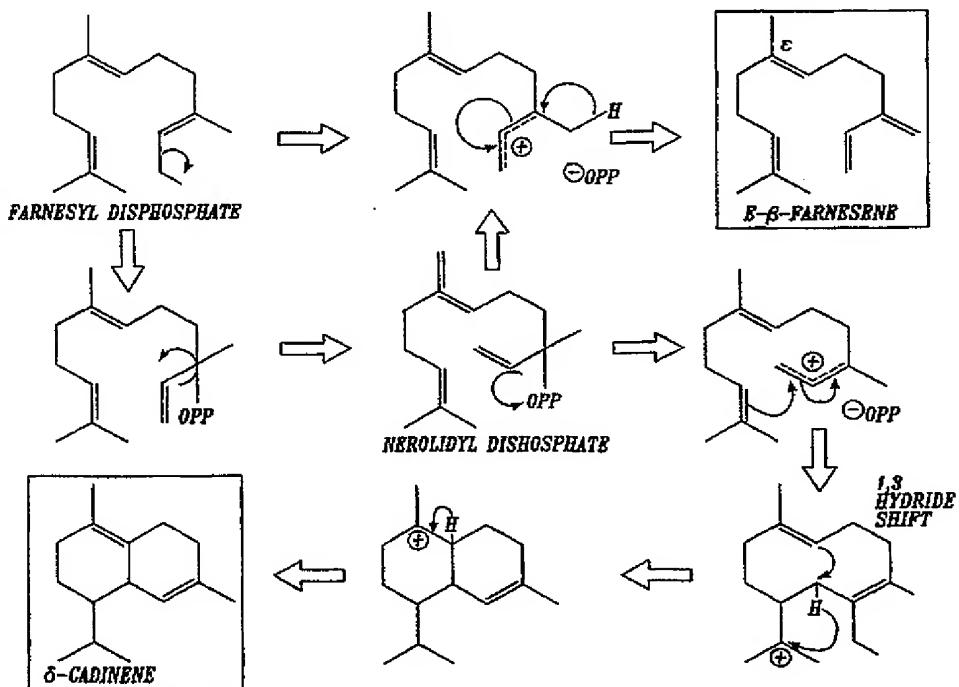
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(54) Title: ISOLATION AND EXPRESSION OF FARNESENE SYNTHASE FROM PEPPERMINT, *MENTHA X PIPERITA*, L.

## (57) Abstract

A cDNA encoding (*E*)- $\beta$ -farnesene synthase from peppermint (*Mentha piperita*) has been isolated and sequenced, and the corresponding amino acid sequence has been determined. Accordingly, an isolated DNA sequence (SEQ ID NO:1) is provided which codes for the expression of (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2), from peppermint (*Mentha piperita*). In other aspects, replicable recombinant cloning vehicles are provided which code for (*E*)- $\beta$ -farnesene synthase, or for base sequence sufficiently complementary to at least a portion of (*E*)- $\beta$ -farnesene synthase DNA or RNA to enable hybridization therewith. In yet other aspects, modified host cells are provided that have been transformed, transfected, infected and/or injected with a recombinant cloning vehicle and/or DNA sequence encoding (*E*)- $\beta$ -farnesene synthase. Thus,



systems and methods are provided for the recombinant expression of the aforementioned recombinant (*E*)- $\beta$ -farnesene synthase that may be used to facilitate its production, isolation and purification in significant amounts. Recombinant (*E*)- $\beta$ -farnesene synthase may be used to obtain expression or enhanced expression of (*E*)- $\beta$ -farnesene synthase in plants in order to enhance the production of (*E*)- $\beta$ -farnesene, or may be otherwise employed for the regulation or expression of (*E*)- $\beta$ -farnesene synthase, or the production of its product.

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**ISOLATION AND EXPRESSION OF FARNESENE SYNTHASE FROM PEPPERMINT, *MENTHA X PIPERITA*, L.**

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5 Hatch Project grant number 0268 from the Agricultural Research Center, Washington  
State University. The government has certain rights in the invention.

**Field of the Invention**

The present invention relates to nucleic acid sequences which code for (*E*)- $\beta$ -farnesene synthases, such as the (*E*)- $\beta$ -farnesene synthase from *Mentha piperita*, and  
10 to vectors containing the sequences, host cells containing the sequences and methods  
of producing recombinant (*E*)- $\beta$ -farnesene synthases and their mutants.

**Background of the Invention**

(*E*)- $\beta$ -farnesene (FIGURE 1) is an acyclic sesquiterpene olefin that occurs in a wide range of both plant and animal taxa. Over 600 papers have been published on  
15 the occurrence of this natural product and its deployment as an important courier in chemical communication. The olefin is found in the essential oil of hundreds of species of both gymnosperms, such as *Torreya taxifolia* (Florida torreya) (Shu, C. K., Lawrence, B. M. and Croom, E. M., Jr. (1995) *J. Essent. Oil Res.* 7, 71-72) and *Larix leptolepis* (larch) (Nabeta, K., Ara, Y., Aoki, Y. and Miyake, M. (1990) *J. Nat. Prod.* 53, 1241-1248), and angiosperms, such as *Robinia pseudoacacia* (black locust) (Kamden, D. P., Gruber, K., Barkman, L. and Gage, D. A. (1994) *J. Essent. Oil Res.* 6, 199-200), *Medicago sativa* (alfalfa) (Kamm, J. A. and Buttery, R. G. (1983) *Entomol. Exp. Appl.* 33, 129-134), *Chamomilla recutita* (chamomile) (Matos, P. J. A., Machiado, M. I. L., Alencar, J. W. and Craveiro, A. A. (1993) *J. Essent. Oil*

- Res. 5, 337-339), *Vitis vinifera* (grapes) (Buchbauer, G., Jirovetz, L., Wasicky, M. and Nikiforov, A. (1994) *J. Essent. Oil Res.* 6, 311-314), *Cannabis sativa* (hemp) (Lemberkovics, E., Veszki, P., Verzar-Petri, G. and Trka, A. (1981) *Sci. Pharm.* 49, 401-408), *Zea mays* (corn) (Turlings, T. C. J., Tumlinson, J. H., Heath, R. R., 5 Proveaux, A. T. and Doolittle, R. E. (1991) *J. Chem. Ecol.* 17, 2235-2251), *Piper nigrum* (black pepper), *Daucus carota* (carrot), and *Mentha x piperita* (peppermint) (Lawrence, B. M. (1972) *Ann. Acad. Bras. Cienc.* 44, (suppl.), 191-197).

While socially dominant male mice produce both  $\alpha$ -farnesene and (*E*)- $\beta$ -farnesene in their urine as pheromones (Novotny, M., Harvey, S. and Jemiolo, B. 10 (1990) *Experientia* 46, 109-113), it is in the insects and plants that the use of (*E*)- $\beta$ -farnesene as a semiochemical is most extensive. (*E*)- $\beta$ -Farnesene is emitted by the Dufour's gland of andrenid bees (Fernandes, A., Duffield, R. M., Wheeler, J. W. and LaBerge, W. E. (1981) *J. Chem. Ecol.* 7, 453-460) and by several genera of ants (Ali, M. F., Morgan, E. D., Attygalle, A. B. and Billen, J. P. J. (1987) *Z. Naturforsch.* 42, 15 955-960; Jackson, B. D., Morgan, E. D. and Billen, J. P. J. (1990) *Naturwiss.* 77, 187-188; Ollert, D. G., Morgan, E. D., Attygalle, A. B. and Billen, J. P. J. (1987) *Z. Naturforsch.* 42, 141-146), where it serves both as a defensive allomone and as a trail pheromone. This sesquiterpene is synthesized *de novo* in the osmeterial glands of larval *Papilio* (Lepidoptera:Papilionidae) as an allomone (Honda, K. (1990) *Insect Biochem.* 20, 245-250), and it functions as a feeding stimulant to the sand fly *Lutzomyia longipalpis* (Diptera:Psychodidae), an important vector of the blood disease leishmaniasis (Tesh, R. B., Guzman, H. and Wilson, M. (1992) *J. Med. Entomol.* 29, 226-231). Several species of predatory carabid beetles use *E*- $\beta$ -farnesene as a prey-finding kairomone (Kiely, J. P., Allen-Williams, L. J., 20 Underwood, N. and Eastwood, E. A. (1996) *J. Insect Behav.* 9, 237-250). When released by corn, this olefin is also a kairomonal oviposition stimulant to the European corn borer (*Ostrinia*) (Binder, B. F., Robbins, J. C. and Wilson, R. L. (1995) *J. Chem. Ecol.* 21, 1315-1327). (*E*)- $\beta$ -farnesene is the major component of pollen odor in *Lupinus* and stimulates pollination behavior in bumblebees (Dobson, H. E. M., 25 Groth, I. and Bergstroem, G. (1996) *Am. J. Bot.* 83, 877-885). Feeding by larval lepidopterans, such as *Heliothis* or *Spodoptera* (Noctuidae), increases the amount of (*E*)- $\beta$ -farnesene released by corn; the volatile olefin is then detected as a synomone by the parasitic wasp *Cotesia marginiventris* (Hymenoptera:Braconidae) for locating the lepidopteran hosts (Turlings, T. C. J., Tumlinson, J. H., Heath, R. R., Proveaux, A. T. 30 and Doolittle, R. E. (1991) *J. Chem. Ecol.* 17, 2235-2251). Circumstantial evidence 35

also suggests the lepidopteran induced production and emission of (*E*)- $\beta$ -farnesene from corn serves as a synomone for *Cotesia kariyai* (Takabayashi, J., Takahashi, S., Dicke, M. and Posthumus, M. A. (1995) *J. Chem. Ecol.* **21**, 273-287) and from cotton leaves as a synomone for *C. marginiventris* (Pare, P. W. and Tumlinson, J. H. (1997) *Nature* **385**, 30-31; Loughrin, J. H., Manukian, A., Heath, R. R., Turlings, T. C. J. and Turnlinson, J. H. (1994) *Proc. Natl. Acad. Sci. USA* **91**, 11836-11840).

Perhaps of greatest significance in plant-insect interactions is the use of (*E*)- $\beta$ -farnesene by most aphid species as an alarm pheromone (Bowers, W. S., Nault, L. R., Webb, R. E. and Dutky, S. R. (1972) *Science* **177**, 1121-1122; Edwards, L. J., 10 Siddall, J. B., Dunham, L. L., Uden, P. and Kislow, C. J. (1973) *Nature* **241**, 126-127). Aphids exposed to (*E*)- $\beta$ -farnesene become agitated and disperse from their host plant (Wohlers, P. (1981) *Z Angew. Entomol.* **92**, 329-336). Alate aphids are usually more sensitive than are apterae species and will often not colonize a host displaying (*E*)- $\beta$ -farnesene. Ants that defend aphids are sensitive to host-emitted (*E*)- $\beta$ -farnesene and, when exposed, will display aggressive behavior (Nault, L. R. and Montgomery, M. E. (1976) *Science* **192**, 1349-1351). (*E*)- $\beta$ -farnesene also mimics the action of juvenile hormone III in some insects (Mauchamp, B. and Pickett, J. J. (1987) *Agronomie* **7**, 523-529), may play a role in control of aphid morphological types, and is acutely toxic to aphids at a dose of 100 ng/aphid (van Oosten, A. M., 15 Gut, J., Harrewijn, P. and Piron, P. G. M. (1990) *Acta Phytopathol. Entomol. Hung.* **25**, 331-342). (*E*)- $\beta$ -farnesene vapor is also toxic to whiteflies (Klijnstra, K. W., Corts, K. A. and van Oosten, A. M. (1992) *Meded. Fac. Landbouwwet.* **57**, 485-491).

Efforts to control aphid behavior by topical application of (*E*)- $\beta$ -farnesene to crops have met with little success, due to volatility and rapid oxidative inactivation in air (Dawson, G. W., Griffiths, D. C., Pickett, J. A., Plumb, R. T., Woodcock, C. M. and Zhang, Z. N. (1988) *Pest. Sci.* **22**, 17-30). Derivatives of (*E*)- $\beta$ -farnesene with reduced volatility, or increased stability, have shown promise in reducing aphid-transmitted viruses, such as barley mosaic virus (Dawson, G. W., Griffiths, D. C., Pickett, J. A., Plumb, R. T., Woodcock, C. M. and Zhang, Z. N. (1988) *Pest. Sci.* **22**, 17-30), potato virus Y (Gibson, R. W., Pickett, J. A., Dawson, G. W., Rice, A. D. and Sibley, M. F. (1984) *Ann. Appl. Entomol.* **104**, 203-209), and beet mosaic virus (Gibson, R. W., Pickett, J. A., Dawson, G. W., Rice, A. D. and Sibley, M. F. (1984) *Ann. Appl. Entomol.* **104**, 203-209). The wild potato *Solanum berthaultii*, which produces (*E*)- $\beta$ -farnesene in type A trichomes, is more repellent to the green peach aphid than are commercial varieties of *S. tuberosum* that produce lower levels of the 30 35

olefin (Gibson, R. W. and Pickett, J. A. (1983) *Nature* **302**, 608-609; Ave, D. A., Gregory, P. and Tingey, W. M. (1987) *Entomol. Exp. App.* **44**, 131-138). In alfalfa, repellency to the blue alfalfa aphid and the pea aphid is correlated with the leaf content of (*E*)- $\beta$ -farnesene, but not with the amount of the co-occurring sesquiterpene 5 caryophyllene (Mostafavi, R., Henning, J. A., Gardea-Torresday, J. and Ray, I. M. (1996) *J. Chem. Ecol.* **22**, 1629-1638).

For plants that produce (*E*)- $\beta$ -farnesene, breeding for increased production has met with some success (Mostafavi, R., Henning, J. A., Gardea-Torresday, J. and Ray, I. M. (1996) *J. Chem. Ecol.* **22**, 1629-1638), but has been limited by genetic 10 variation in these species. (*E*)- $\beta$ -farnesene synthase has been purified from maritime pine (*Pinus pinaster*) and characterized (Salin, F., Pauly, G., Charon, J. and Gleizes, M. (1995) *J. Plant Phys.* **146**, 203-209), but the gene has not yet been isolated from any source. A cDNA clone for (*E*)- $\beta$ -farnesene synthase would, by transgenic manipulation, provide a valuable addition to the arsenal of natural compounds active 15 in host plant resistance. The substrate for (*E*)- $\beta$ -farnesene synthase is farnesyl diphosphate, a ubiquitous isoprenoid intermediate involved in cytoplasmic phytosterol biosynthesis. Sesquiterpene synthases lack plastidial targeting sequences and are localized to the cytoplasm (Chappell, J. (1995) *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **46**, 521-547). Therefore, even in plants that do not normally produce 20 sesquiterpenes, a recombinant (*E*)- $\beta$ -farnesene synthase would be directed to the cytoplasm where substrate is supplied by the mevalonate pathway and where production of (*E*)- $\beta$ -farnesene should result.

#### Summary of the Invention

In accordance with the foregoing, a cDNA encoding (*E*)- $\beta$ -farnesene synthase 25 from peppermint (*Mentha piperita*) has been isolated and sequenced, and the corresponding amino acid sequence has been deduced. Accordingly, the present invention relates to isolated DNA sequences which code for the expression of (*E*)- $\beta$ -farnesene synthase, such as the sequence designated SEQ ID NO:1 which encodes an (*E*)- $\beta$ -farnesene synthase protein (SEQ ID NO:2) from peppermint (*Mentha piperita*). 30 Additionally, the present invention relates to isolated, recombinant (*E*)- $\beta$ -farnesene synthase proteins from peppermint (*Mentha piperita*). In other aspects, the present invention is directed to replicable recombinant cloning vehicles comprising a nucleic acid sequence, e.g., a DNA sequence which codes for an (*E*)- $\beta$ -farnesene synthase, or 35 for a base sequence sufficiently complementary to at least a portion of DNA or RNA encoding (*E*)- $\beta$ -farnesene synthase to enable hybridization therewith (e.g., antisense

RNA or fragments of DNA complementary to a portion of DNA or RNA molecules encoding (*E*)- $\beta$ -farnesene synthase which are useful as polymerase chain reaction primers or as probes for (*E*)- $\beta$ -farnesene synthase or related genes). In yet other aspects of the invention, modified host cells are provided that have been transformed, 5 transfected, infected and/or injected with a recombinant cloning vehicle and/or DNA sequence of the invention. Thus, the present invention provides for the recombinant expression of (*E*)- $\beta$ -farnesene synthase, and the inventive concepts may be used to facilitate the production, isolation and purification of significant quantities of recombinant (*E*)- $\beta$ -farnesene synthase (or of its primary enzyme products) for 10 subsequent use, to obtain expression or enhanced expression of (*E*)- $\beta$ -farnesene synthase in plants, microorganisms or animals, or may be otherwise employed in an environment where the regulation or expression of (*E*)- $\beta$ -farnesene synthase is desired for the production of this synthase, or its enzyme product, or derivatives thereof.

Brief Description of the Drawings

15 The foregoing aspects and many of the attendant advantages of this invention will become more readily appreciated as the same becomes better understood by reference to the following detailed description, when taken in conjunction with the accompanying drawings, wherein:

20 FIGURE 1. The sesquiterpene synthase substrate, farnesyl diphosphate, and sesquiterpene olefins found in peppermint essential oil.

FIGURE 2. Radio-GC of the sesquiterpene olefins generated from [1-<sup>3</sup>H]farnesyl diphosphate by an enzyme preparation from peppermint oil gland secretory cells. The olefin fraction of steam-distilled peppermint oil was used as internal standard, and only the portion of the chromatogram containing the 25 sesquiterpene olefins is shown.

FIGURE 3A. GC-MS of the products generated from farnesyl diphosphate by the recombinant (*E*)- $\beta$ -farnesene synthase. Panel A: Total ion chromatogram. Numbered peaks are sesquiterpene olefins.

30 FIGURE 3B. Mass spectrum and retention time of peak 1 designated in FIGURE 3 A.

FIGURE 3C. Mass spectrum and retention time of authentic (*E*)- $\beta$ -farnesene from parley oil.

35 FIGURE 3D. Mass spectrum and retention time of peak 6 designated in FIGURE 3 A. The spectrum of this minor product is compromised by the low ion abundance and the corresponding prominence of background ions.

FIGURE 3E. Mass spectrum and retention time of authentic  $\delta$ -cadinene.

FIGURE 4. Proposed mechanism for the formation of (*E*)- $\beta$ -farnesene and  $\delta$ -cadinene from farnesyl diphosphate. OPP denotes the diphosphate moiety. Ionization of the enzyme-bound nerolidyl diphosphate intermediate and proton elimination can also produce (*E*)- $\beta$ -farnesene.

FIGURE 5. Monoterpene olefins generated from the alternate substrate geranyl diphosphate by recombinant (*E*)- $\beta$ -farnesene synthase.

Detailed Description of the Preferred Embodiment

As used herein, the terms "amino acid" and "amino acids" refer to all naturally occurring L- $\alpha$ -amino acids or their residues. The amino acids are identified by either the single-letter or three-letter designations:

	Asp	D	aspartic acid	Ile	I	isoleucine
	Thr	T	threonine	Leu	L	leucine
	Ser	S	serine	Tyr	Y	tyrosine
15	Glu	E	glutamic acid	Phe	F	phenylalanine
	Pro	P	proline	His	H	histidine
	Gly	G	glycine	Lys	K	lysine
	Ala	A	alanine	Arg	R	arginine
	Cys	C	cysteine	Trp	W	tryptophan
20	Val	V	valine	Gln	Q	glutamine
	Met	M	methionine	Asn	N	asparagine

As used herein, the term "nucleotide" means a monomeric unit of DNA or RNA containing a sugar moiety (pentose), a phosphate and a nitrogenous heterocyclic base. The base is linked to the sugar moiety via the glycosidic carbon (1' carbon of pentose) and that combination of base and sugar is called a nucleoside. The base characterizes the nucleotide with the four bases of DNA being adenine ("A"), guanine ("G"), cytosine ("C") and thymine ("T"). Inosine ("I") is a synthetic base that can be used to substitute for any of the four, naturally-occurring bases (A, C, G or T). The four RNA bases are A, G, C and uracil ("U"). The nucleotide sequences described herein comprise a linear array of nucleotides connected by phosphodiester bonds between the 3' and 5' carbons of adjacent pentoses.

"Oligonucleotide" refers to short length single or double stranded sequences of deoxyribonucleotides linked via phosphodiester bonds. The oligonucleotides are chemically synthesized by known methods and purified, for example, on polyacrylamide gels.

The term "(E)- $\beta$ -farnesene synthase" refers to an enzyme that is capable of converting farnesyl diphosphate to (E)- $\beta$ -farnesene.

The term "essential oil plant," or "essential oil plants," refers to a group of plant species that produce high levels of monoterpenoid and/or sesquiterpenoid and/or diterpenoid oils, and/or high levels of monoterpenoid and/or sesquiterpenoid and/or diterpenoid resins. The foregoing oils and/or resins account for greater than about 0.005% of the fresh weight of an essential oil plant that produces them. The essential oils and/or resins are more fully described, for example, in E. Guenther, *The Essential Oils*, Vols. I-VI, R.E. Krieger Publishing Co., Huntington N.Y., 1975, incorporated herein by reference. The essential oil plants include, but are not limited to:

Lamiaceae, including, but not limited to, the following species: Ocimum (basil), Lavandula (Lavender), Origanum (oregano), Mentha (mint), Salvia (sage), Rosmecinus (rosemary), Thymus (thyme), Satureja and Monarda.

Umbelliferae, including, but not limited to, the following species: Carum (caraway), Anethum (dill), feniculum (fennel) and Daucus (carrot).

Asteraceae (Compositae), including, but not limited to, the following species: Artemisia (tarragon, sage brush), Tanacetum (tansy).

Rutaceae (e.g., citrus plants); Rosaceae (e.g., roses); Myrtaceae (e.g., eucalyptus, Melaleuca); the Gramineae (e.g., Cymbopogon (citronella)); Geranaceae (Geranium) and certain conifers including Abies (e.g., Canadian balsam), Cedrus (cedar) and Thuja and Juniperus.

The range of essential oil plants is more fully set forth in *E. Guenther, The Essential Oils, Vols. I-VI, R.E. Krieger Publishing Co., Huntington N.Y., 1975*, which is incorporated herein by reference.

The term "angiosperm" refers to a class of plants that produce seeds that are enclosed in an ovary.

The term "gymnosperm" refers to a class of plants that produce seeds that are not enclosed in an ovary.

Abbreviations used are: bp, base pairs; dpm, disintegrations per minute; DTT, 30 dithiothreitol; EDTA, ethylenediaminetetraacetic acid; FDP, farnesyl diphosphate; GC, gas chromatography; GDP, geranyl diphosphate; GGDP, geranylgeranyl diphosphate; I, identity; IPTG, isopropyl- $\beta$ -D-thiogalactopyranoside; LB, Luria-Bertani; Mopso, 3-(*N*-morpholino)-2-hydroxypropane-sulfonic acid; MS, mass spectrometry; PVPP, polyvinylpolypyrrolidone; S, similarity.

The term "percent identity" (%I) means the percentage of amino acids or nucleotides that occupy the same relative position when two amino acid sequences, or two nucleic acid sequences, are aligned side by side.

5       The term "percent similarity" (%S) is a statistical measure of the degree of relatedness of two compared protein sequences. The percent similarity is calculated by a computer program that assigns a numerical value to each compared pair of amino acids based on chemical similarity (e.g., whether the compared amino acids are acidic, basic, hydrophobic, aromatic, etc.) and/or evolutionary distance as measured by the minimum number of base pair changes that would be required to convert a codon 10 encoding one member of a pair of compared amino acids to a codon encoding the other member of the pair. Calculations are made after a best fit alignment of the two sequences has been made empirically by iterative comparison of all possible alignments. (Henikoff, S. and Henikoff, J.G., *Proc. Nat'l Acad Sci USA* **89**: 10915-10919, 1992).

15       The abbreviation "SSC" refers to a buffer used in nucleic acid hybridization solutions. One liter of the 20X (twenty times concentrate) stock SSC buffer solution (pH 7.0) contains 175.3 g sodium chloride and 88.2 g sodium citrate.

20       The terms "alteration", "amino acid sequence alteration", "variant" and "amino acid sequence variant" refer to (E)- $\beta$ -farnesene synthase molecules with some differences in their amino acid sequences as compared to the corresponding, native, i.e., naturally-occurring, (E)- $\beta$ -farnesene synthases. Ordinarily, the variants will possess at least about 70% homology with the corresponding native (E)- $\beta$ -farnesene synthases, and preferably, they will be at least about 80% homologous with the corresponding, native (E)- $\beta$ -farnesene synthases. The amino acid sequence variants 25 of the (E)- $\beta$ -farnesene synthases falling within this invention possess substitutions, deletions, and/or insertions at certain positions. Sequence variants of (E)- $\beta$ -farnesene synthases may be used to attain desired enhanced or reduced enzymatic activity, modified regiochemistry or stereochemistry, or altered substrate utilization or product distribution.

30       Substitutional (E)- $\beta$ -farnesene synthase variants are those that have at least one amino acid residue in the native (E)- $\beta$ -farnesene synthase sequence removed and a different amino acid inserted in its place at the same position. The substitutions may be single, where only one amino acid in the molecule has been substituted, or they may be multiple, where two or more amino acids have been substituted in the same 35 molecule. Substantial changes in the activity of the (E)- $\beta$ -farnesene synthase

molecules of the present invention may be obtained by substituting an amino acid with a side chain that is significantly different in charge and/or structure from that of the native amino acid. This type of substitution would be expected to affect the structure of the polypeptide backbone and/or the charge or hydrophobicity of the molecule in 5 the area of the substitution.

Moderate changes in the activity of the (E)- $\beta$ -farnesene synthase molecules of the present invention would be expected by substituting an amino acid with a side chain that is similar in charge and/or structure to that of the native molecule. This type of substitution, referred to as a conservative substitution, would not be expected 10 to substantially alter either the structure of the polypeptide backbone or the charge or hydrophobicity of the molecule in the area of the substitution.

Insertional (E)- $\beta$ -farnesene synthase variants are those with one or more amino acids inserted immediately adjacent to an amino acid at a particular position in the native (E)- $\beta$ -farnesene synthase molecule. Immediately adjacent to an amino acid 15 means connected to either the  $\alpha$ -carboxy or  $\alpha$ -amino functional group of the amino acid. The insertion may be one or more amino acids. Ordinarily, the insertion will consist of one or two conservative amino acids. Amino acids similar in charge and/or structure to the amino acids adjacent to the site of insertion are defined as conservative. Alternatively, this invention includes insertion of an amino acid with a 20 charge and/or structure that is substantially different from the amino acids adjacent to the site of insertion.

Deletional variants are those where one or more amino acids in the native (E)- $\beta$ -farnesene synthase molecules have been removed. Ordinarily, deletional variants will have one or two amino acids deleted in a particular region of the (E)- $\beta$ -farnesene 25 synthase molecule.

The terms "biological activity", "biologically active", "activity" and "active" refer to the ability of the (E)- $\beta$ -farnesene synthases of the present invention to catalyze the formation of (E)- $\beta$ -farnesene from farnesyl diphosphate. (E)- $\beta$ -farnesene synthase activity is measured in an enzyme activity assay, such as the assay described 30 in Example 1 herein. Amino acid sequence variants of the (E)- $\beta$ -farnesene synthases of the present invention may have desirable altered biological activity including, for example, altered reaction kinetics, substrate utilization, product distribution or other characteristics such as regiochemistry and stereochemistry.

The terms "DNA sequence encoding", "DNA encoding" and "nucleic acid 35 encoding" refer to the order or sequence of deoxyribonucleotides along a strand of

deoxyribonucleic acid. The order of these deoxyribonucleotides determines the order of amino acids along the translated polypeptide chain. The DNA sequence thus codes for the amino acid sequence.

The terms "replicable expression vector" and "expression vector" refer to a piece of DNA, usually double-stranded, which may have inserted into it another piece of DNA (the insert DNA) such as, but not limited to, a cDNA molecule. The vector is used to transport the insert DNA into a suitable host cell. The insert DNA may be derived from the host cell, or may be derived from a different cell or organism. Once in the host cell, the vector can replicate independently of or coincidental with the host chromosomal DNA, and several copies of the vector and its inserted DNA may be generated. In addition, the vector contains the necessary elements that permit translating the insert DNA into a polypeptide. Many molecules of the polypeptide encoded by the insert DNA can thus be rapidly synthesized.

The terms "transformed host cell," "transformed" and "transformation" refer to the introduction of DNA into a cell. The cell is termed a "host cell", and it may be a prokaryotic or a eukaryotic cell. Typical prokaryotic host cells include various strains of *E. coli*. Typical eukaryotic host cells are plant cells, such as maize cells, yeast cells, insect cells or animal cells. The introduced DNA is usually in the form of a vector containing an inserted piece of DNA. The introduced DNA sequence may be from the same species as the host cell or from a different species from the host cell, or it may be a hybrid DNA sequence, containing some foreign DNA and some DNA derived from the host species.

In accordance with the present invention, a cDNA (SEQ ID NO:1) encoding (E)- $\beta$ -farnesene synthase (SEQ ID NO:2) from peppermint (*Mentha piperita*) was isolated and sequenced in the following manner. An enriched cDNA library was constructed from peppermint secretory cell clusters consisting of the eight glandular cells subtending the oil droplet. These cell clusters were harvested by leaf surface abrasion and the RNA contained therein was isolated. mRNA was purified by oligo-dT cellulose chromatography, and 5  $\mu$ g of mRNA was used to construct a  $\lambda$ ZAPII cDNA library.

Plasmids were excised from the library *en masse* and used to transform *E. coli* strain XLORL. Approximately 150 individual plasmid-bearing strains were grown in 5 ml LB media overnight, and the corresponding plasmids were purified before partial 5'-sequencing. Putative terpenoid synthase genes were identified by sequence comparison using the BLAST program of the GCG Wisconsin Package ver. 8.

Bluescript plasmids harboring unique full-length cDNA inserts with high similarity to known plant terpenoid synthases were tested for functional expression following transformation into *E. coli* XL1-Blue cells. A single extract, from the bacteria containing clone p43, including the cDNA insert set forth in SEQ ID NO:1, produced 5 a sesquiterpene olefin from [1-<sup>3</sup>H]FDP, and this clone was selected for further study.

A cell-free extract of *E. coli* XL-1 Blue cells harboring the plasmid p43, including the cDNA insert set forth in SEQ ID NO:1, was prepared and shown to be capable of catalyzing the divalent metal ion-dependent conversion of [1-<sup>3</sup>H]FDP to labeled sesquiterpene olefins. Control reactions, employing extracts of XL1-Blue 10 cells transformed with pBluescript lacking the insert, evidenced no detectable production of sesquiterpene olefins from [1-<sup>3</sup>H]FDP, thereby demonstrating that a cDNA clone (SEQ ID NO:1) encoding (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) had been acquired.

The recombinant (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) was inactive with 15 the C<sub>20</sub> substrate analog [1-<sup>3</sup>H]GGDP, but was able to catalyze the divalent cation-dependent conversion of the C<sub>10</sub> analog [1-<sup>3</sup>H]GDP to monoterpane olefins. Control reactions, employing extracts of XL1-Blue cells transformed with pBluescript lacking the insert, evidenced no detectable production of monoterpane olefins from [1-<sup>3</sup>H]GDP, thereby confirming that the monoterpane synthase activity expressed 20 from the cDNA insert of p43 (SEQ ID NO:1) was a function of the (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2). This is the first report describing the utilization of GDP by a sesquiterpene synthase.

Complete sequencing of the (*E*)- $\beta$ -farnesene synthase cDNA (SEQ ID NO:1) contained in p43 revealed an insert size of 1959 bp encoding an open reading frame of 25 550 amino acids with a deduced molecular weight of 63,829. The deduced amino acid sequence of the (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) lacks a plastidial targeting peptide. Like all other known terpenoid synthases, (*E*)- $\beta$ -farnesene synthase is rich in tryptophan (1.8%) and arginine (5.5%) residues, and bears a DDXXD motif 30 (SEQ ID NO:3) (residues 301-305 of SEQ ID NO:2) which is believed to coordinate the divalent metal ion chelated to the substrate diphosphate group. The enzyme has a deduced isoelectric point at pH 5.16.

The isolation of a cDNA (SEQ ID NO:1) encoding (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) permits the development of efficient expression systems for this functional enzyme; provides useful tools for examining the developmental regulation 35 of (*E*)- $\beta$ -farnesene synthase; permits investigation of the reaction mechanism(s) of this

enzyme, and permits the isolation of other (*E*)- $\beta$ -farnesene synthases. The isolation of an (*E*)- $\beta$ -farnesene synthase cDNA (SEQ ID NO:1) also permits the transformation of a wide range of organisms in order to enhance, enable or otherwise alter, the synthesis of (*E*)- $\beta$ -farnesene.

5        Although the (*E*)- $\beta$ -farnesene synthase protein set forth in SEQ ID NO:2 lacks a plastidial targeting sequence, a targeting sequence from another protein can be included in the (*E*)- $\beta$ -farnesene synthase amino terminus. Transport sequences well known in the art (See, for example, the following publications, the cited portions of which are incorporated by reference herein: von Heijne et al., *Eur. J. Biochem.*,  
10      180:535-545, 1989; Stryer, *Biochemistry*, W.H. Freeman and Company, New York, NY, p. 769 [1988]) may be employed to direct (*E*)- $\beta$ -farnesene synthase to other cellular or extracellular locations.

15      In addition to the native (*E*)- $\beta$ -farnesene synthase amino acid sequence of SEQ ID NO:2, sequence variants produced by deletions, substitutions, mutations and/or insertions are intended to be within the scope of the invention except insofar as limited by the prior art. The (*E*)- $\beta$ -farnesene synthase amino acid sequence variants of this invention may be constructed by mutating the DNA sequences that encode the wild-type synthases, such as by using techniques commonly referred to as site-directed mutagenesis. Nucleic acid molecules encoding the (*E*)- $\beta$ -farnesene synthases 20      of the present invention can be mutated by a variety of PCR techniques well known to one of ordinary skill in the art. (See, for example, the following publications, the cited portions of which are incorporated by reference herein: "PCR Strategies", M.A. Innis, D.H. Gelfand and J.J. Sninsky, eds., 1995, Academic Press, San Diego, CA (Chapter 14); "PCR Protocols: A Guide to Methods and Applications", M.A. Innis, 25      D.H. Gelfand, J.J. Sninsky and T.J. White, eds., Academic Press, NY (1990).

By way of non-limiting example, the two primer system utilized in the Transformer Site-Directed Mutagenesis kit from Clontech, may be employed for introducing site-directed mutants into the (*E*)- $\beta$ -farnesene synthase genes of the present invention. Following denaturation of the target plasmid in this system, two 30      primers are simultaneously annealed to the plasmid; one of these primers contains the desired site-directed mutation, the other contains a mutation at another point in the plasmid resulting in elimination of a unique restriction site. Second strand synthesis is then carried out, tightly linking these two mutations, and the resulting plasmids are transformed into a *mutS* strain of *E. coli*. Plasmid DNA is isolated from the 35      transformed bacteria, restricted with the relevant restriction enzyme (thereby

linearizing the unmutated plasmids), and then retransformed into *E. coli*. This system allows for generation of mutations directly in an expression plasmid, without the necessity of subcloning or generation of single-stranded phagemids. The tight linkage of the two mutations and the subsequent linearization of unmutated plasmids results in 5 high mutation efficiency and allows minimal screening. Following synthesis of the initial restriction site primer, this method requires the use of only one new primer type per mutation site. Rather than prepare each positional mutant separately, a set of "designed degenerate" oligonucleotide primers can be synthesized in order to introduce all of the desired mutations at a given site simultaneously. Transformants 10 can be screened by sequencing the plasmid DNA through the mutagenized region to identify and sort mutant clones. Each mutant DNA can then be fully sequenced or restricted and analyzed by electrophoresis on Mutation Detection Enhancement gel (J.T. Baker) to confirm that no other alterations in the sequence have occurred (by band shift comparison to the unmutagenized control).

15 Again, by way of non-limiting example, the two primer system utilized in the QuikChange™ Site-Directed Mutagenesis kit from Stratagene (LaJolla, California), may be employed for introducing site-directed mutants into the (*E*)- $\beta$ -farnesene synthase genes of the present invention. Double-stranded plasmid DNA, containing the insert bearing the target mutation site, is denatured and mixed with two 20 oligonucleotides complementary to each of the strands of the plasmid DNA at the target mutation site. The annealed oligonucleotide primers are extended using *Pfu* DNA polymerase, thereby generating a mutated plasmid containing staggered nicks. After temperature cycling, the unmutated, parental DNA template is digested with restriction enzyme *DpnI* which cleaves methylated or hemimethylated DNA, but 25 which does not cleave unmethylated DNA. The parental, template DNA is almost always methylated or hemimethylated since most strains of *E.coli*, from which the template DNA is obtained, contain the required methylase activity. The remaining, annealed vector DNA incorporating the desired mutation(s) is transformed into *E. coli*.

30 The mutated (*E*)- $\beta$ -farnesene synthase gene can be cloned into a pET (or other) overexpression vector that can be employed to transform *E. coli* such as strain *E. coli* BL21(DE3)pLysS, for high level production of the mutant protein, and purification by standard protocols. Examples of plasmid vectors and *E. coli* strains that can be used to express high levels of the (*E*)- $\beta$ -farnesene synthase proteins of the 35 present invention are set forth in Sambrook et al, *Molecular Cloning, A Laboratory*

*Manual*, 2nd Edition (1989), Chapter 17. The method of FAB-MS mapping can be employed to rapidly check the fidelity of mutant expression. This technique provides for sequencing segments throughout the whole protein and provides the necessary confidence in the sequence assignment. In a mapping experiment of this type, protein 5 is digested with a protease (the choice will depend on the specific region to be modified since this segment is of prime interest and the remaining map should be identical to the map of unmutagenized protein). The set of cleavage fragments is fractionated by microbore HPLC (reversed phase or ion exchange, again depending on the specific region to be modified) to provide several peptides in each fraction, and 10 the molecular weights of the peptides are determined by FAB-MS. The masses are then compared to the molecular weights of peptides expected from the digestion of the predicted sequence, and the correctness of the sequence quickly ascertained. Since the exemplary mutagenesis techniques set forth herein produce site-directed mutations, sequencing of the altered peptide should not be necessary if the mass 15 spectrograph agrees with prediction. If necessary to verify a changed residue, CAD-tandem MS/MS can be employed to sequence the peptides of the mixture in question, or the target peptide can be purified for subtractive Edman degradation or carboxypeptidase Y digestion depending on the location of the modification.

In the design of a particular site directed mutagenesis experiment, it is 20 generally desirable to first make a non-conservative substitution (e.g., Ala for Cys, His or Glu) and determine if activity is greatly impaired as a consequence. The properties of the mutagenized protein are then examined with particular attention to the kinetic parameters of  $K_m$  and  $k_{cat}$  as sensitive indicators of altered function, from which changes in binding and/or catalysis *per se* may be deduced by comparison to the 25 native enzyme. If the residue is by this means demonstrated to be important by activity impairment, or knockout, then conservative substitutions can be made, such as Asp for Glu to alter side chain length, Ser for Cys, or Arg for His. For hydrophobic segments, it is largely size that is usefully altered, although aromatics can also be substituted for alkyl side chains. Changes in the normal product distribution 30 can indicate which step(s) of the reaction sequence have been altered by the mutation. Modification of the hydrophobic pocket can be employed to change binding conformations for substrates and result in altered regiochemistry and/or stereochemistry.

Other site directed mutagenesis techniques may also be employed with the 35 nucleotide sequences of the invention. For example, restriction endonuclease

digestion of DNA followed by ligation may be used to generate deletion variants of *(E)*- $\beta$ -farnesene synthase, as described in section 15.3 of Sambrook et al. *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Press, New York, NY [1989], incorporated herein by reference. A similar strategy may be 5 used to construct insertion variants, as described in section 15.3 of Sambrook et al., *supra*.

Oligonucleotide-directed mutagenesis may also be employed for preparing substitution variants of this invention. It may also be used to conveniently prepare the deletion and insertion variants of this invention. This technique is well known in the 10 art as described by Adelman et al. (*DNA* 2:183 [1983]); Sambrook et al., *supra*; "Current Protocols in Molecular Biology", 1991, Wiley (NY), F.T. Ausubel, R. Brent, R.E. Kingston, D.D. Moore, J.D. Seidman, J.A. Smith and K. Struhi, eds, incorporated herein by reference.

Generally, oligonucleotides of at least 25 nucleotides in length are used to 15 insert, delete or substitute two or more nucleotides in the *(E)*- $\beta$ -farnesene synthase molecule. An optimal oligonucleotide will have 12 to 15 perfectly matched nucleotides on either side of the nucleotides coding for the mutation. To mutagenize wild-type *(E)*- $\beta$ -farnesene synthase, the oligonucleotide is annealed to the single-stranded DNA template molecule under suitable hybridization conditions. A DNA 20 polymerizing enzyme, usually the Klenow fragment of *E. coli* DNA polymerase I, is then added. This enzyme uses the oligonucleotide as a primer to complete the synthesis of the mutation-bearing strand of DNA. Thus, a heteroduplex molecule is formed such that one strand of DNA encodes the wild-type synthase inserted in the vector, and the second strand of DNA encodes the mutated form of the synthase 25 inserted into the same vector. This heteroduplex molecule is then transformed into a suitable host cell.

Mutants with more than one amino acid substituted may be generated in one 30 of several ways. If the amino acids are located close together in the polypeptide chain, they may be mutated simultaneously using one oligonucleotide that codes for all of the desired amino acid substitutions. If, however, the amino acids are located some distance from each other (separated by more than ten amino acids, for example) it is more difficult to generate a single oligonucleotide that encodes all of the desired changes. Instead, one of two alternative methods may be employed. In the first method, a separate oligonucleotide is generated for each amino acid to be substituted. 35 The oligonucleotides are then annealed to the single-stranded template DNA

simultaneously, and the second strand of DNA that is synthesized from the template will encode all of the desired amino acid substitutions. An alternative method involves two or more rounds of mutagenesis to produce the desired mutant. The first round is as described for the single mutants: wild-type (*E*)- $\beta$ -farnesene synthase DNA 5 is used for the template, an oligonucleotide encoding the first desired amino acid substitution(s) is annealed to this template, and the heteroduplex DNA molecule is then generated. The second round of mutagenesis utilizes the mutated DNA produced in the first round of mutagenesis as the template. Thus, this template already contains one or more mutations. The oligonucleotide encoding the additional 10 desired amino acid substitution(s) is then annealed to this template, and the resulting strand of DNA now encodes mutations from both the first and second rounds of mutagenesis. This resultant DNA can be used as a template in a third round of mutagenesis, and so on.

A gene encoding (*E*)- $\beta$ -farnesene synthase may be incorporated into any 15 organism (intact plant, animal, microbe, etc.), or cell culture derived therefrom, that produces substrates that can be converted to (*E*)- $\beta$ -farnesene. An (*E*)- $\beta$ -farnesene synthase gene may be introduced into any organism for a variety of purposes including, but not limited to: production of (*E*)- $\beta$ -farnesene synthase, or its product (*E*)- $\beta$ -farnesene; enhancement of the rate of production and/or the absolute amount of 20 (*E*)- $\beta$ -farnesene; enhancement of protection of plants against pests and pathogens, for example by producing (*E*)- $\beta$ -farnesene to act as a pollinator attractant synomone for predators and parasites of plant pests, or as an aphid alarm pheromone. While the nucleic acid molecules of the present invention can be introduced into any organism, the nucleic acid molecules of the present invention will preferably be introduced into a 25 plant species.

Eukaryotic expression systems may be utilized for the production of (*E*)- $\beta$ -farnesene synthase since they are capable of carrying out any required posttranslational modifications and of directing the enzyme to the proper cellular compartment. A representative eukaryotic expression system for this purpose uses 30 the recombinant baculovirus, *Autographa californica* nuclear polyhedrosis virus (AcNPV; M.D. Summers and G.E. Smith, *A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures* [1986]; Luckow et al., *Bio-technology*, 6:47-55 [1987]) for expression of the (*E*)- $\beta$ -farnesene synthases of the invention. Infection of insect cells (such as cells of the species *Spodoptera frugiperda*) with the 35 recombinant baculoviruses allows for the production of large amounts of the (*E*)- $\beta$ -

- farnesene synthase proteins. In addition, the baculovirus system has other important advantages for the production of recombinant (*E*)- $\beta$ -farnesene synthase. For example, baculoviruses do not infect humans and can therefore be safely handled in large quantities. In the baculovirus system, a DNA construct is prepared including a DNA segment encoding (*E*)- $\beta$ -farnesene synthase and a vector. The vector may comprise the polyhedron gene promoter region of a baculovirus, the baculovirus flanking sequences necessary for proper cross-over during recombination (the flanking sequences comprise about 200-300 base pairs adjacent to the promoter sequence) and a bacterial origin of replication which permits the construct to replicate in bacteria.
- 5      The vector is constructed so that (i) the DNA segment is placed adjacent (or operably linked or "downstream" or "under the control of") to the polyhedron gene promoter and (ii) the promoter/(*E*)- $\beta$ -farnesene synthase combination is flanked on both sides by 200-300 base pairs of baculovirus DNA (the flanking sequences).
- 10     To produce the (*E*)- $\beta$ -farnesene synthase DNA construct, a cDNA clone

15     encoding the full length (*E*)- $\beta$ -farnesene synthase is obtained using methods such as those described herein. The DNA construct is contacted in a host cell with baculovirus DNA of an appropriate baculovirus (that is, of the same species of baculovirus as the promoter encoded in the construct) under conditions such that recombination is effected. The resulting recombinant baculoviruses encode the full

20     (*E*)- $\beta$ -farnesene synthase. For example, an insect host cell can be cotransfected or transfected separately with the DNA construct and a functional baculovirus. Resulting recombinant baculoviruses can then be isolated and used to infect cells to effect production of the (*E*)- $\beta$ -farnesene synthase. Host insect cells include, for example, *Spodoptera frugiperda* cells, that are capable of producing a baculovirus-expressed (*E*)- $\beta$ -farnesene synthase. Insect host cells infected with a recombinant baculovirus of the present invention are then cultured under conditions allowing expression of the baculovirus-encoded (*E*)- $\beta$ -farnesene synthase. (*E*)- $\beta$ -farnesene synthase thus produced is then extracted from the cells using methods known in the art.

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30     Other eukaryotic microbes such as yeasts may also be used to practice this invention. The baker's yeast *Saccharomyces cerevisiae*, is a commonly used yeast, although several other strains are available. The plasmid YRp7 (Stinchcomb et al., *Nature*, 282:39 [1979]; Kingsman et al., *Gene* 7:141 [1979]; Tschemper et al., *Gene*, 10:157 [1980]) is commonly used as an expression vector in *Saccharomyces*. This plasmid contains the trp1 gene that provides a selection marker for a mutant strain of

yeast lacking the ability to grow in the absence of tryptophan, such as strains ATCC No. 44,076 and PEP4-1 (Jones, *Genetics*, 85:12 [1977]). The presence of the *trp1* lesion as a characteristic of the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan.

5 Yeast host cells are generally transformed using the polyethylene glycol method, as described by Hinnen (*Proc. Natl. Acad. Sci. USA*, 75:1929 [1978]). Additional yeast transformation protocols are set forth in Gietz et al., *N.A.R.*, 20(17):1425(1992); Reeves et al., *FEMS*, 99(2-3):193-197, (1992), both of which publications are incorporated herein by reference.

10 Suitable promoting sequences in yeast vectors include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., *J. Biol. Chem.*, 255:2073 [1980]) or other glycolytic enzymes (Hess et al., *J. Adv. Enzyme Reg.* 7:149 [1968]; Holland et al., *Biochemistry*, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase,  
15 glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase. In the construction of suitable expression plasmids, the termination sequences associated with these genes are also ligated into the expression vector 3' of the sequence desired to be expressed to provide polyadenylation of the mRNA and termination. Other  
20 promoters that have the additional advantage of transcription controlled by growth conditions are the promoter region for alcohol dehydrogenase 2, isocytchrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, and the aforementioned glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Any plasmid vector containing yeast-compatible  
25 promoter, origin of replication and termination sequences is suitable.

Cell cultures derived from multicellular organisms, such as plants, may be used as hosts to practice this invention. Transgenic plants can be obtained, for example, by transferring plasmids that encode (*E*)- $\beta$ -farnesene synthase and a selectable marker gene, e.g., the kan gene encoding resistance to kanamycin, into *Agrobacterium tumifaciens* containing a helper Ti plasmid as described in Hoeckema et al., *Nature*, 303:179-181 [1983] and culturing the *Agrobacterium* cells with leaf slices, or other tissues or cells, of the plant to be transformed as described by An et al., *Plant Physiology*, 81:301-305 [1986]. Transformation of cultured plant host cells is normally accomplished through *Agrobacterium tumifaciens*. Cultures of mammalian host cells and other host cells that do not have rigid cell membrane barriers are usually

transformed using the calcium phosphate method as originally described by Graham and Van der Eb (*Virology*, 52:546 [1978]) and modified as described in sections 16.32-16.37 of Sambrook et al., *supra*. However, other methods for introducing DNA into cells such as Polybrene (Kawai and Nishizawa, *Mol. Cell. Biol.*, 4:1172 [1984]), protoplast fusion (Schaffner, *Proc. Natl. Acad. Sci. USA*, 77:2163 [1980]), electroporation (Neumann et al., *EMBO J.*, 1:841 [1982]), and direct microinjection into nuclei (Capecchi, *Cell*, 22:479 [1980]) may also be used. Additionally, animal transformation strategies are reviewed in Monastersky G.M. and Robl, J.M., *Strategies in Transgenic Animal Science*, ASM Press, Washington, D.C., 1995, incorporated herein by reference. Transformed plant calli may be selected through the selectable marker by growing the cells on a medium containing, e.g., kanamycin, and appropriate amounts of phytohormone such as naphthalene acetic acid and benzyladenine for callus and shoot induction. The plant cells may then be regenerated and the resulting plants transferred to soil using techniques well known to those skilled in the art.

In addition, a gene regulating (*E*)- $\beta$ -farnesene synthase production can be incorporated into the plant along with a necessary promoter which is inducible. In the practice of this embodiment of the invention, a promoter that only responds to a specific external or internal stimulus is fused to the target cDNA. Thus, the gene will not be transcribed except in response to the specific stimulus. As long as the gene is not being transcribed, its gene product and enzyme product are not produced.

An illustrative example of a responsive promoter system that can be used in the practice of this invention is the glutathione-S-transferase (GST) system in maize. GSTs are a family of enzymes that can detoxify a number of hydrophobic electrophilic compounds that often are used as pre-emergent herbicides (Weigand et al., *Plant Molecular Biology*, 7:235-243 [1986]). Studies have shown that the GSTs are directly involved in causing this enhanced herbicide tolerance. This action is primarily mediated through a specific 1.1 kb mRNA transcription product. In short, maize has a naturally occurring quiescent gene already present that can respond to external stimuli and that can be induced to produce a gene product. This gene has previously been identified and cloned. Thus, in one embodiment of this invention, the promoter is removed from the GST responsive gene and attached to an (*E*)- $\beta$ -farnesene synthase gene that previously has had its native promoter removed. This engineered gene is the combination of a promoter that responds to an external chemical stimulus and a gene responsible for successful production of (*E*)- $\beta$ -farnesene synthase.

In addition to the methods described above, several methods are known in the art for transferring cloned DNA into a wide variety of plant species, including gymnosperms, angiosperms, monocots and dicots (see, e.g., Glick and Thompson, eds., *Methods in Plant Molecular Biology*, CRC Press, Boca Raton, Florida [1993], incorporated by reference herein). Representative examples include electroporation-facilitated DNA uptake by protoplasts in which an electrical pulse transiently permeabilizes cell membranes, permitting the uptake of a variety of biological molecules, including recombinant DNA (Rhodes et al., *Science*, 240:204-207 [1988]); treatment of protoplasts with polyethylene glycol (Lyznik et al., 5 *Plant Molecular Biology*, 13:151-161 [1989]); and bombardment of cells with DNA-laden microprojectiles which are propelled by explosive force or compressed gas to 10 penetrate the cell wall (Klein et al., *Plant Physiol.* 91:440-444 [1989] and Boynton et al., *Science*, 240:1534-1538 [1988]). Transformation of *Taxus* species can be achieved, for example, by employing the methods set forth in Han et al, *Plant* 15 *Science*, 95:187-196 (1994), incorporated by reference herein. A method that has been applied to Rye plants (*Secale cereale*) is to directly inject plasmid DNA, including a selectable marker gene, into developing floral tillers (de la Pena et al., *Nature* 325:274-276 (1987)). Further, plant viruses can be used as vectors to transfer genes to plant cells. Examples of plant viruses that can be used as vectors to 20 transform plants include the Cauliflower Mosaic Virus (Brisson et al., *Nature* 310: 511-514 (1984); Additionally, plant transformation strategies and techniques are reviewed in Birch, R.G., *Ann Rev Plant Phys Plant Mol Biol*, 48:297 (1997); Forester et al., *Exp. Agric.*, 33:15-33 (1997). The aforementioned publications disclosing plant transformation techniques are incorporated herein by reference, and 25 minor variations make these technologies applicable to a broad range of plant species.

Each of these techniques has advantages and disadvantages. In each of the techniques, DNA from a plasmid is genetically engineered such that it contains not only the gene of interest, but also selectable and screenable marker genes. A selectable marker gene is used to select only those cells that have integrated copies of 30 the plasmid (the construction is such that the gene of interest and the selectable and screenable genes are transferred as a unit). The screenable gene provides another check for the successful culturing of only those cells carrying the genes of interest. A commonly used selectable marker gene is neomycin phosphotransferase II (NPT II). This gene conveys resistance to kanamycin, a compound that can be added directly to 35 the growth media on which the cells grow. Plant cells are normally susceptible to

kanamycin and, as a result, die. The presence of the NPT II gene overcomes the effects of the kanamycin and each cell with this gene remains viable. Another selectable marker gene which can be employed in the practice of this invention is the gene which confers resistance to the herbicide glufosinate (Basta). A screenable gene 5 commonly used is the  $\beta$ -glucuronidase gene (GUS). The presence of this gene is characterized using a histochemical reaction in which a sample of putatively transformed cells is treated with a GUS assay solution. After an appropriate incubation, the cells containing the GUS gene turn blue.

The plasmid containing one or more of these genes is introduced into either 10 plant protoplasts or callus cells by any of the previously mentioned techniques. If the marker gene is a selectable gene, only those cells that have incorporated the DNA package survive under selection with the appropriate phytotoxic agent. Once the appropriate cells are identified and propagated, plants are regenerated. Progeny from the transformed plants must be tested to insure that the DNA package has been 15 successfully integrated into the plant genome.

Mammalian host cells may also be used in the practice of the invention. Examples of suitable mammalian cell lines include monkey kidney CVI line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line 293S (Graham et al., *J. Gen. Virol.*, 36:59 [1977]); baby hamster kidney cells (BHK, 20 ATCC CCL 10); Chinese hamster ovary cells (Urlab and Chasin, *Proc. Natl. Acad. Sci USA* 77:4216 [1980]); mouse sertoli cells (TM4, Mather, *Biol. Reprod.*, 23:243 [1980]); monkey kidney cells (CVL-76, ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells 25 (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor cells (MMT 060562, ATCC CCL 51); rat hepatoma cells (HTC, ML54, Baumann et al., *J. Cell Biol.*, 85:1 [1980]); and TRI cells (Mather et al., *Annals N.Y. Acad. Sci.*, 383:44 [1982]). Expression vectors for these cells ordinarily include (if necessary) DNA sequences for 30 an origin of replication, a promoter located in front of the gene to be expressed, a ribosome binding site, an RNA splice site, a polyadenylation site, and a transcription terminator site.

Promoters used in mammalian expression vectors are often of viral origin. These viral promoters are commonly derived from polyoma virus, Adenovirus 2, and 35 most frequently Simian Virus 40 (SV40). The SV40 virus contains two promoters

that are termed the early and late promoters. These promoters are particularly useful because they are both easily obtained from the virus as one DNA fragment that also contains the viral origin of replication (Fiers et al., *Nature*, 273:113 [1978]). Smaller or larger SV40 DNA fragments may also be used, provided they contain the 5 approximately 250-bp sequence extending from the HindIII site toward the BglII site located in the viral origin of replication.

Alternatively, promoters that are naturally associated with the foreign gene (homologous promoters) may be used provided that they are compatible with the host cell line selected for transformation.

10 An origin of replication may be obtained from an exogenous source, such as SV40 or other virus (e.g., Polyoma, Adeno, VSV, BPV) and inserted into the cloning vector. Alternatively, the origin of replication may be provided by the host cell chromosomal replication mechanism. If the vector containing the foreign gene is integrated into the host cell chromosome, the latter is often sufficient.

15 The use of a secondary DNA coding sequence can enhance production levels of (*E*)- $\beta$ -farnesene synthase in transformed cell lines. The secondary coding sequence typically comprises the enzyme dihydrofolate reductase (DHFR). The wild-type form of DHFR is normally inhibited by the chemical methotrexate (MTX). The level of DHFR expression in a cell will vary depending on the amount of MTX added to the 20 cultured host cells. An additional feature of DHFR that makes it particularly useful as a secondary sequence is that it can be used as a selection marker to identify transformed cells. Two forms of DHFR are available for use as secondary sequences, wild-type DHFR and MTX-resistant DHFR. The type of DHFR used in a particular host cell depends on whether the host cell is DHFR deficient (such that it either 25 produces very low levels of DHFR endogenously, or it does not produce functional DHFR at all). DHFR-deficient cell lines such as the CHO cell line described by Urlaub and Chasin, *supra*, are transformed with wild-type DHFR coding sequences. After transformation, these DHFR-deficient cell lines express functional DHFR and are capable of growing in a culture medium lacking the nutrients hypoxanthine, 30 glycine and thymidine. Nontransformed cells will not survive in this medium.

The MTX-resistant form of DHFR can be used as a means of selecting for transformed host cells in those host cells that endogenously produce normal amounts of functional DHFR that is MTX sensitive. The CHO-K1 cell line (ATCC No. CL 61) possesses these characteristics, and is thus a useful cell line for this purpose. The 35 addition of MTX to the cell culture medium will permit only those cells transformed

with the DNA encoding the MTX-resistant DHFR to grow. The nontransformed cells will be unable to survive in this medium.

Prokaryotes may also be used as host cells for the initial cloning steps of this invention. They are particularly useful for rapid production of large amounts of DNA, 5 for production of single-stranded DNA templates used for site-directed mutagenesis, for screening many mutants simultaneously, and for DNA sequencing of the mutants generated. Suitable prokaryotic host cells include *E. coli* K12 strain 94 (ATCC No. 31,446), *E. coli* strain W3110 (ATCC No. 27,325) *E. coli* X1776 (ATCC No. 31,537), and *E. coli* B; however many other strains of *E. coli*, such as HB101, 10 JM101, NM522, NM538, NM539, and many other species and genera of prokaryotes including bacilli such as *Bacillus subtilis*, other enterobacteriaceae such as *Salmonella typhimurium* or *Serratia marcesans*, and various *Pseudomonas* species may all be used as hosts. Prokaryotic host cells or other host cells with rigid cell walls are preferably transformed using the calcium chloride method as described in section 1.82 15 of Sambrook et al., *supra*. Alternatively, electroporation may be used for transformation of these cells. Prokaryote transformation techniques are set forth in Dower, W.J., in Genetic Engineering, Principles and Methods, 12:275-296, Plenum Publishing Corp., 1990; Hanahan et al., *Meth. Enzymol.*, 204:63 (1991).

As a representative example, cDNA sequences encoding (*E*)- $\beta$ -farnesene 20 synthase may be transferred to the (His)<sub>6</sub>\*Tag pET vector commercially available (from Novagen) for overexpression in *E. coli* as heterologous host. This pET expression plasmid has several advantages in high level heterologous expression systems. The desired cDNA insert is ligated in frame to plasmid vector sequences encoding six histidines followed by a highly specific protease recognition site 25 (thrombin) that are joined to the amino terminus codon of the target protein. The histidine "block" of the expressed fusion protein promotes very tight binding to immobilized metal ions and permits rapid purification of the recombinant protein by immobilized metal ion affinity chromatography. The histidine leader sequence is then cleaved at the specific proteolysis site by treatment of the purified protein with 30 thrombin, and the (*E*)- $\beta$ -farnesene synthase again purified by immobilized metal ion affinity chromatography, this time using a shallower imidazole gradient to elute the recombinant synthases while leaving the histidine block still adsorbed. This overexpression-purification system has high capacity, excellent resolving power and is fast, and the chance of a contaminating *E. coli* protein exhibiting similar binding 35 behavior (before and after thrombin proteolysis) is extremely small.

As will be apparent to those skilled in the art, any plasmid vectors containing replicon and control sequences that are derived from species compatible with the host cell may also be used in the practice of the invention. The vector usually has a replication site, marker genes that provide phenotypic selection in transformed cells,  
5 one or more promoters, and a polylinker region containing several restriction sites for insertion of foreign DNA. Plasmids typically used for transformation of *E. coli* include pBR322, pUC18, pUC19, pUC118, pUC119, and Bluescript M13, all of which are described in sections 1.12-1.20 of Sambrook et al., *supra*. However, many other suitable vectors are available as well. These vectors contain genes coding for  
10 ampicillin and/or tetracycline resistance which enables cells transformed with these vectors to grow in the presence of these antibiotics.

The promoters most commonly used in prokaryotic vectors include the  $\beta$ -lactamase (penicillinase) and lactose promoter systems (Chang et al., *Nature*, 375:615 [1978]; Itakura et al., *Science*, 198:1056 [1977]; Goeddel et al., *Nature*,  
15 281:544 [1979]) and a tryptophan (*trp*) promoter system (Goeddel et al., *Nucl. Acids Res.*, 8:4057 [1980]; EPO Appl. Publ. No. 36,776), and the alkaline phosphatase systems. While these are the most commonly used, other microbial promoters have been utilized, and details concerning their nucleotide sequences have been published,  
20 enabling a skilled worker to ligate them functionally into plasmid vectors (see Siebenlist et al., *Cell*, 20:269 [1980]).

Many eukaryotic proteins normally secreted from the cell contain an endogenous secretion signal sequence as part of the amino acid sequence. Thus, proteins normally found in the cytoplasm can be targeted for secretion by linking a signal sequence to the protein. This is readily accomplished by ligating DNA encoding a signal sequence to the 5' end of the DNA encoding the protein and then expressing this fusion protein in an appropriate host cell. The DNA encoding the signal sequence may be obtained as a restriction fragment from any gene encoding a protein with a signal sequence. Thus, prokaryotic, yeast, and eukaryotic signal sequences may be used herein, depending on the type of host cell utilized to practice  
25 the invention. The DNA and amino acid sequence encoding the signal sequence portion of several eukaryotic genes including, for example, human growth hormone, proinsulin, and proalbumin are known (see Stryer, *Biochemistry* W.H. Freeman and Company, New York, NY, p. 769 [1988]), and can be used as signal sequences in appropriate eukaryotic host cells. Yeast signal sequences, as for example acid  
30 phosphatase (Arima et al., *Nuc. Acids Res.*, 11:1657 [1983]),  $\alpha$ -factor, alkaline  
35 phosphatase (Arima et al., *Nuc. Acids Res.*, 11:1657 [1983]),  $\alpha$ -factor, alkaline

phosphatase and invertase may be used to direct secretion from yeast host cells. Prokaryotic signal sequences from genes encoding, for example, LamB or OmpF (Wong et al., *Gene*, 68:193 [1988]), MalE, PhoA, or beta-lactamase, as well as other genes, may be used to target proteins from prokaryotic cells into the culture medium.

5 Trafficking sequences from plants, animals and microbes can be employed in the practice of the invention to direct the (*E*)- $\beta$ -farnesene synthase proteins of the present invention to the cytoplasm, endoplasmic reticulum, mitochondria or other cellular components, or to target the protein for export to the medium. These considerations apply to the overexpression of (*E*)- $\beta$ -farnesene synthase, and to  
10 direction of expression within cells or intact organisms to permit gene product function in any desired location.

The construction of suitable vectors containing DNA encoding replication sequences, regulatory sequences, phenotypic selection genes and the (*E*)- $\beta$ -farnesene synthase DNA of interest are prepared using standard recombinant DNA procedures.  
15 Isolated plasmids and DNA fragments are cleaved, tailored, and ligated together in a specific order to generate the desired vectors, as is well known in the art (see, for example, Sambrook et al., *supra*).

As discussed above, (*E*)- $\beta$ -farnesene synthase variants are preferably produced by means of mutation(s) that are generated using the method of site-specific  
20 mutagenesis. This method requires the synthesis and use of specific oligonucleotides that encode both the sequence of the desired mutation and a sufficient number of adjacent nucleotides to allow the oligonucleotide to stably hybridize to the DNA template.

The foregoing may be more fully understood in connection with the following  
25 representative examples, in which "Plasmids" are designated by a lower case p followed by an alphanumeric designation. The starting plasmids used in this invention are either commercially available, publicly available on an unrestricted basis, or can be constructed from such available plasmids using published procedures. In addition, other equivalent plasmids are known in the art and will be apparent to the ordinary  
30 artisan.

"Digestion", "cutting" or "cleaving" of DNA refers to catalytic cleavage of the DNA with an enzyme that acts only at particular locations in the DNA. These enzymes are called restriction endonucleases, and the site along the DNA sequence where each enzyme cleaves is called a restriction site. The restriction enzymes used in  
35 this invention are commercially available and are used according to the instructions

supplied by the manufacturers. (See also sections 1.60-1.61 and sections 3.38-3.39 of Sambrook et al., *supra*.)

"Recovery" or "isolation" of a given fragment of DNA from a restriction digest means separation of the resulting DNA fragment on a polyacrylamide or an agarose gel by electrophoresis, identification of the fragment of interest by comparison of its mobility versus that of marker DNA fragments of known molecular weight, removal of the gel section containing the desired fragment, and separation of the gel from DNA. This procedure is known generally. For example, see Lawn et al. (*Nucleic Acids Res.*, 9:6103-6114 [1982]), and Goeddel et al. (*Nucleic Acids Res.*, 10 *supra*).

The following examples merely illustrate the best mode now contemplated for practicing the invention, but should not be construed to limit the invention.

Example 1

Essential Oil Analysis and Cell-Free Assay

15 **Plant Material and Reagents.** Unless stated otherwise, the following plant materials and reagents were used in the experiments reported in this and succeeding Examples. *Mentha x piperita* L. cv. 'Black Mitcham' was propagated from rhizomes as previously described (Gershenson, J., McCaskill, D., Rajaonarivony, J. I. M., Mihaliak, C., Karp, F. and Croteau, R. (1992) *Anal. Biochem.* 200, 130-138). The 20 preparations of [ $1\text{-}^3\text{H}$ ]geranyl diphosphate (GDP) (250 Ci/mol), [ $1\text{-}^3\text{H}$ ]farnesyl diphosphate (FDP) (125 Ci/mol), and [ $1\text{-}^3\text{H}$ ]geranylgeranyl diphosphate (GGDP) (118 Ci/mol) have been previously reported (Croteau, R., Alonso, W. R., Koepp, A. E. and Johnson, M. A. (1994) *Arch. Biochem. Biophys.* 309, 184-192; Dixit, V. M., Laskovics, F. M., Noall, W. I. and Poulter, C. D. (1981) *J. Org. Chem.* 46, 25 1967-1969; LaFever, R. E., StoferVogel, B. and Croteau, R. (1994) *Arch. Biochem. Biophys.* 313, 139-149). Terpenoid standards were from our own collection or were prepared from plant material purchased locally.  $\alpha$ -Farnesene was a gift from Dr. J. Brown (Washington State University),  $\delta$ -cadinene was a gift from Dr. M. Essenberg (Oklahoma State University), and commercially steam distilled peppermint oil was a 30 gift from I. P. Callison and Sons, Inc., Chehalis, WA. All other biochemicals and reagents were purchased from Sigma Chemical Co. or Aldrich Chemical Co., unless otherwise noted.

35 **Sesquiterpene Analysis.** Unless stated otherwise, the following procedure was utilized to analyze sesquiterpene content and composition in the experiments reported in this and succeeding Examples. Young, mature peppermint leaves were

harvested and hydrodistilled from NH<sub>4</sub>HCO<sub>3</sub>-buffered water with simultaneous pentane extraction (Maarse, H. and Kepner, R. E. (1970) *J. Agr. Chem.* **18**, 1095-1101). The organic phase was passed through a column of MgSO<sub>4</sub>-silica gel (Mallinckrodt SilicAR-60) to provide the olefin fraction for GC-MS analysis.

- 5      Authentic (*E*)- $\beta$ -farnesene was prepared by pentane extraction (followed by silica gel fractionation) of macerated ginger (*Zingiber officinale*) root, black pepper oleoresin (*Piper nigrum*), bergamot oil (*Citrus bergamot*), parsley oil (*Petroselinum crispum*), or field-grown (Yakima Valley, WA) commercial peppermint oil (Lawrence, B. M. (1972) *Ann. Acad. Bras. Cienc.* **44**, (suppl.), 191-197); all of these sources are  
10     reported to contain (*E*)- $\beta$ -farnesene.

**Instrumental Analysis.** The following instrumentation was utilized in this Example and all succeeding Examples, unless stated otherwise. Radio-GC was performed on a Gow-Mac 550P instrument (He carrier 40 ml/min, injector 220°C, detector 250°C and 150 mA) attached to a Packard 894 gas proportional counter.

- 15     The column (3.18 mm i.d. by 3.66 m stainless steel with 15% polyethylene glycol ester (AT1000 Alltech) on Gas Chrom Q was programmed from 150°C (5 min. hold) to 220°C at 5°C/min. Thermal conductivity and radioactivity outputs were monitored after calibration with an external radiochemical standard, and ~20,000 dpm of tritiated product was injected with data analysis using Turbochrome Navigator ver. 4.1  
20     software (Perkin-Elmer). Liquid scintillation counting was performed in toluene:ethanol (70:30, v/v) containing 0.4% Omnifluor (DuPont NEN) using a Packard 460 CD spectrometer (<sup>3</sup>H efficiency ~43%). GC-MS analysis employed a Hewlett-Packard 6890-5972 system with a 5MS capillary column (0.25 mm i.d. by 30 m with 0.25 Tm coating of 5% phenyl methyl siloxane). Injections were made  
25     cool on-column at 40°C with oven programming from 40°C (50°C/min) to 50°C (5 min hold), then 10°C/min to 250°C, then 50°C/min to 300°C. Separations were made under a constant flow of 0.7 ml He/min. Mass spectral data were collected at 70 eV and analyzed using Hewlett-Packard Chemstation software.

- Cell-Free Assays. Peppermint oil gland secretory cells were isolated from immature leaves as previously described (Gershenson, J., McCaskill, D., Rajaonarivony, J. I. M., Mihaliak, C., Karp, F. and Croteau, R. (1992) *Anal. Biochem.* **200**, 130-138, incorporated herein by reference) and sonically disrupted (Braun-Sonic 2000 microprobe at maximum power for three 30-second bursts with 30-second chilling period at 0-4°C between bursts) into assay buffer consisting of  
30     25 mM Mopso (pH 7.0), 10 mM sodium ascorbate, 25 mM KCl, 10 mM DTT and  
35     25 mM DTT and

10% glycerol, and supplemented with 0.5% (w/v) PVPP and 1% (w/v) Amberlite XAD-4 polystyrene resin. The sonicate was centrifuged at 3700 x g for 15 minutes, and an aliquot of the supernatant was then placed in a 10 ml screw-capped glass test tube containing divalent metal ions (10 mM MgCl<sub>2</sub> and 1 mM MnCl<sub>2</sub>) and substrate 5 (7.3 µM [1-<sup>3</sup>H]FDP). The aqueous layer was overlaid with 1 ml pentane and the sealed tube was incubated at 30°C for two hours. The pentane overlay was then collected and the aqueous layer was extracted twice (1 ml) with pentane. The combined pentane extracts were passed through an anhydrous MgSO<sub>4</sub>-silica gel 10 column to obtain the labeled hydrocarbon fraction for GC-MS analysis, or for radio-GC analysis using peppermint oil as an internal standard.

**Essential Oil Analysis.** To assess the probable abundance of (*E*)-β-farnesene synthase in peppermint gland secretory cells, the exclusive site of essential oil biosynthesis (Gershenzon, J., McCaskill, D., Rajaonarivony, J. I. M., Mihaliak, C., Karp, F. and Croteau, R. (1992) *Anal. Biochem.* **200**, 130-138), the sesquiterpene 15 olefin fraction of field-distilled peppermint oil was analyzed by GC-MS and shown to contain β-caryophyllene (39%), γ-cadinene (33%), β-bourbonene (11%), (*E*)-β-farnesene (2.9%), δ-cadinene (2.0%), germacrene D (1.3%), copaene (1.3%) and α-humulene (1.2%) (FIGURE 1), as well as several other minor components (<1% each). GC-MS analysis of the oil distilled from greenhouse material revealed a similar 20 composition, except that the amount of γ-cadinene was higher (53%), β-bourbonene was conspicuously absent, and the (*E*)-β-farnesene content was 3.4%. Although (*E*)-β-farnesene was not one of the more prominent sesquiterpenes of peppermint, the abundance was sufficient to suggest that cloning of the corresponding synthase by random sequencing of an enriched, oil gland cDNA library might be possible.

25       **Cell-free extracts.** To gain a preliminary assessment of the target activity, cell-free extracts of peppermint oil gland secretory cells (Gershenzon, J., McCaskill, D., Rajaonarivony, J. I. M., Mihaliak, C., Karp, F. and Croteau, R. (1992) *Anal. Biochem.* **200**, 130-138), were assayed for the divalent metal ion-dependent conversion of [1-<sup>3</sup>H]farnesyl diphosphate to sesquiterpene olefins (Cane, D. E. (1990) *Chem. Rev.* **90**, 1089-1103). Radio-GC analysis of the derived biosynthetic products 30 (FIGURE 2) revealed the presence of two major components identified as caryophyllene and γ-cadinene. However, the separation of the labeled olefins was insufficient to resolve (*E*)-β-farnesene from caryophyllene, or δ-cadinene-from γ-cadinene. Both of these minor components appear at the trailing edges of the major 35 peaks but are, nevertheless, coincident with the authentic standards, indicating the

corresponding biosynthetic capability. No  $\beta$ -bourbonene was synthesized from FDP by this system.

**Example 2**

**Cloning and Expression in *E.coli* of a cDNA Encoding (*E*)- $\beta$ -Farnesene Synthase (SEQ ID NO:1)**

5

**Library Construction and Clone Identification.** Initial cloning of full-length terpenoid biosynthetic genes from the peppermint oil gland cDNA library was successful and established a very high degree of enrichment for these target sequences. For example, the monoterpene cyclase, limonene synthase (Colby, S. M., Alonso, W. R., Katahira, E. J., McGarvey, D. J. and Croteau, R. (1993) *J. Biol. Chem.* **268**, 23016-23024), represents approximately 4% of the library. This fact, plus the availability of automated sequencing capability, led to the possibility of randomly sequencing the library in search of cDNA species encoding other terpenoid synthases, including the (*E*)- $\beta$ -farnesene synthase which was shown to be operational in this plant by both sesquiterpene analysis and cell-free assay.

An enriched cDNA library was constructed from peppermint secretory cell clusters consisting of the eight glandular cells subtending the oil droplet. These cell clusters were harvested by a leaf surface abrasion technique (Gershenson, J., McCaskill, D., Rajaonarivony, J. I. M., Mihaliak, C., Karp, F. and Croteau, R. (1992) *Anal. Biochem.* **200**, 130-138), and the RNA contained therein was isolated using the protocol of Logemann et al. (Logemann, J., Schell, J. and Willmitzer, L. (1987) *Anal. Biochem.* **163**, 16-20). mRNA was purified by oligo-dT cellulose chromatography (Pharmacia), and 5  $\mu$ g of mRNA was used to construct a  $\lambda$ ZAPII cDNA library according to the manufacturer's instructions (Stratagene).

25

Plasmids were excised from the library *en masse* and used to transform *E. coli* strain XLOR as per the manufacturer's instructions (Stratagene). Approximately 150 individual plasmid-bearing strains were grown in 5 ml LB media overnight, and the corresponding plasmids were purified using a Qiawell 8 Ultraplasmid Kit (Qiagen) before partial 5'-sequencing by the Dye-Deoxy™ method using an ABI Sequenator at the Laboratory for Biotechnology and Bioanalysis at Washington State University. Putative terpenoid synthase genes were identified by sequence comparison using the BLAST program of the GCG Wisconsin Package ver. 8. Bluescript plasmids harboring unique full-length cDNA inserts with high similarity to known plant terpenoid synthases were tested for functional expression following transformation into *E. coli* XL1-Blue cells. A single extract, from the bacteria containing clone p43,

including the cDNA insert sequence set forth in SEQ ID NO:1, produced a sesquiterpene olefin from [1-<sup>3</sup>H]FDP, and this clone was selected for further study.

**Bacterial Expression and Characterization of (*E*)- $\beta$ -Farnesene Synthase (SEQ ID NO:2).** *E. coli* XL1-Blue harboring p43 (including the cDNA insert sequence set forth in SEQ ID NO:1), or empty pBluescript plasmid as a control, were grown overnight at 37°C in LB medium containing 100  $\mu$ g ampicillin/ml. A 50  $\mu$ l aliquot of the overnight culture was used to inoculate 5 ml of fresh LB medium, and the culture was grown at 37°C with vigorous agitation to  $A_{600}$  0.5 before induction with 1 mM IPTG. After an additional two hours of growth, the suspension was centrifuged (1000  $\times$  g, 15 min, 4°C), the media removed, and the pelleted cells resuspended in 1 ml of cold assay buffer containing 1 mM EDTA. The cells were disrupted by sonication with a microprobe as previously described, except that only two 20-second bursts were employed. The chilled sonicate was cleared by centrifugation and the supernatant was assayed for sesquiterpene synthase activity as before, or for monoterpene synthase activity (with 4.5  $\mu$ M [1-<sup>3</sup>H]GDP) or diterpene synthase activity (with 10  $\mu$ M [1-<sup>3</sup>H]GGDP). In all cases, the pentane-soluble reaction products were purified by MgSO<sub>4</sub>-silica gel chromatography, as above, to prepare the olefin fraction for further analysis.

A cell-free extract of *E. coli* XL-1 Blue cells harboring the plasmid p43 (including the cDNA insert sequence set forth in SEQ ID NO:1) was prepared and shown to be capable of catalyzing the divalent metal ion-dependent conversion of [1-<sup>3</sup>H]FDP to labeled sesquiterpene olefins. Radio-GC analysis (data not shown) and GC-MS analysis (FIGURE 3) of this sesquiterpene olefin fraction demonstrated that the major biosynthetic product (85%) was (*E*)- $\beta$ -farnesene by matching of both retention time and mass spectrum to those of the authentic standard obtained from several natural sources. Lesser amounts of (*Z*)- $\beta$ -farnesene (8%) and  $\delta$ -cadinene (5%), as well as three other minor products (less than 1% each; all seemingly of the cadinene-type based on MS), were also produced. Control reactions, employing extracts of XL1-Blue cells transformed with pBluescript lacking the cDNA insert having the sequence set forth in SEQ ID NO:1, evidenced no detectable production of sesquiterpene olefins from [1-<sup>3</sup>H]FDP, thereby demonstrating that a cDNA clone encoding (*E*)- $\beta$ -farnesene synthase had been acquired.

Multiple product formation is a common feature of the terpenoid synthases, and may be a consequence of the electrophilic reaction mechanism catalyzed by these enzymes in which highly reactive carbocationic intermediates are generated (Cane,

D. E. (1990) *Chem. Rev.* **90**, 1089-1103; Croteau, R. (1987) *Chem. Rev.* **87**, 929-954). (*E*)- $\beta$ -farnesene is one of the simplest sesquiterpene olefins that can be derived from FDP, in a reaction involving divalent metal ion-assisted ionization of the diphosphate ester and deprotonation from the C-3 methyl of the resulting carbocation (FIGURE 4). The formation of  $\delta$ -cadinene (FIGURE 4) involves a considerably more extended reaction sequence, in which a preliminary isomerization step (to nerolidyl diphosphate) is required to permit the ionization-dependent cyclization to the macrocycle, followed by 1,3-hydride shift, closure of the second ring, and deprotonation to the bicyclic product. The small amount of  $\delta$ -cadinene produced by the recombinant synthase (SEQ ID NO:2) from FDP is interesting in light of the abundance of this bicyclic olefin in the sesquiterpene fraction of peppermint oil and the efficient production of this olefin in oil gland extracts; these observations suggest that an additional and distinct  $\delta$ -cadinene synthase must operate in peppermint.

The recombinant (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) was inactive with the C<sub>20</sub> substrate analog [1-<sup>3</sup>H]GGDP, but was able to catalyze the divalent cation-dependent conversion of the C<sub>10</sub> analog [1-<sup>3</sup>H]GDP to monoterpene olefins. Although the rate of conversion of GDP to these products was less than 3% of the rate of conversion of FDP to sesquiterpene olefins at saturation, a more diverse spectrum of products was formed (see FIGURE 5 for structures). The cyclic monoterpenes limonene (48%) and terpinolene (15%), and the acyclic monoterpene analog of  $\beta$ -farnesene, myrcene (15%), were the most abundant products as determined by both radio-GC and GC-MS analysis (data not shown). Lesser amounts of  $\gamma$ -terpinene (7%), (*Z*)-ocimene (6%), (*E*)-ocimene (7%), and sabinene (3%) were also observed as products. Control reactions, employing extracts of XL1-Blue cells transformed with pBluescript lacking the insert, evidenced no detectable production of monoterpene olefins from [1-<sup>3</sup>H]GDP, thereby confirming that the monoterpene synthase activity expressed from p43 was a function of the (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2). This is the first report describing the utilization of GDP by a sesquiterpene synthase. Because monoterpene biosynthesis is localized to plastids, as is diterpene biosynthesis, whereas sesquiterpene biosynthesis occurs in the cytoplasm (Chappell, J. (1995) *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **46**, 521-547), the utilization of GDP as a substrate by (*E*)- $\beta$ -farnesene synthase is unlikely to be of physiological relevance and may simply reflect the lack of evolutionary pressure to discern the chain length of this isoprenoid substrate to which the enzyme is not exposed *in vivo*.

Example 3Sequence Analysis of the p43 cDNA Insert (SEQ ID NO:1)

Complete sequencing of the (*E*)- $\beta$ -farnesene synthase cDNA (SEQ ID NO:1) contained in p43 revealed an insert size of 1959 bp encoding an open reading frame of 550 amino acids with a deduced molecular weight of 63,829. A putative starting methionine codon was identified which was out of frame with the vector  $\beta$ -galactosidase starting methionine; however, a fortuitous stop codon in the 5'-untranslated region, 46 bp upstream of the synthase translation start site and in frame with the  $\beta$ -galactosidase fusion sequence, allowed polycistronic translation of the 10 cDNA free of vector-derived sequence. The deduced amino acid sequence of the (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) lacks a plastidial targeting peptide (Keegstra, K., Olsen, J. J. and Theg, S. M. (1989) *Ann. Rev. Plant Physiol. Plant Mol. Biol.* **40**, 471-501), typical of monoterpene and diterpene synthases (Colby, S. M., Alonso, W. R., Katahira, E. J., McGarvey, D. J. and Croteau, R. (1993) *J. Biol. Chem.* **268**, 15 23016-23024; Stofer Vogel, B., Wildung, M. R., Vogel, G. and Croteau, R. (1996) *J. Biol. Chem.* **271**, 23262-23268; Wildung, M. R. and Croteau, R. (1996) *J. Biol. Chem.* **271**, 9201-9204), but consistent with all known plant-derived sesquiterpene synthases (Fachinni, P. J. and Chappell, J. (1992) *Proc. Natl. Acad. Sci. USA* **89**, 20 11088-11092; Back, K. and Chappell, J. (1995) *J. Biol. Chem.* **270**, 7375-7381; Chen, X. Y., Chen, Y., Heinstein, P. and Davisson, V. J. (1996) *Arch. Biochem. Biophys.* **324**, 255-266) which are directed to the cytoplasm. Like all other known terpenoid synthases, (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) is rich in tryptophan (1.8%) and arginine (5.5%) residues, and bears a DDXXD motif (residues 301-305)(SEQ ID NO:3) which is believed to coordinate the divalent metal ion chelated to 25 the substrate diphosphate group (Marrero, O. F., Poultier, C. D. and Edwards, P. A. (1992) *J. Biol. Chem.* **267**, 21873-21878); the enzyme (SEQ ID NO:2) has a deduced isoelectric point at pH 5.16.

The deduced amino acid sequence of the farnesene synthase (SEQ ID NO:2) is most similar to that of the sesquiterpene cyclase *epi*-aristolochene synthase from 30 tobacco (Fachinni, P. J. and Chappell, J. (1992) *Proc. Natl. Acad. Sci. USA* **89**, 11088-11092) in exhibiting 62% similarity (S) and 49% identity (I). This peppermint synthase (SEQ ID NO:2) also closely resembles the three other known angiosperm sesquiterpene cyclases (vetispiradiene synthase from *Hyoscyamus muticus* (Back, K. and Chappell, J. (1995) *J. Biol. Chem.* **270**, 7375-7381) at 63% S and 40% I,  $\delta$ -cadinene synthase from cotton (Chen, X. Y., Chen, Y., Heinstein, P. and Davisson,

V. J. (1996) *Arch. Biochem. Biophys.* **324**, 255-266) at 60% S and 37% I, and germacrene C synthase from tomato at 57% S and 34% I (unpublished), and also the diterpene cyclase, casbene synthase (Mau, C. J. D. and West, C. A. (1994) *Proc. Natl. Acad. Sci. USA* **91**, 8497-8501), from castor bean (at 61 % S and 35% I). Since 5 (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) produces a small amount of  $\delta$ -cadinene, but cannot be the major source of  $\delta$ -cadinene in peppermint, it is tempting to speculate that the farnesene synthase (SEQ ID NO:2) represents either a progenitor, or an altered form of cadinene synthase in which the ability to catalyze the more complex bicyclization reaction has been lost.

Surprisingly, (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) is no more closely related to monoterpene synthases from the Lamiaceae (limonene synthase from spearmint (Colby, S.M., Alonso, W. R., Katahira, E. J., McGarvey, D. J. and Croteau, R. (1993) *J. Biol. Chem.* **268**, 23016-23024) with 51% S and 30% I; sabinene synthase and 1,8-cineole synthase from culinary sage with 50% S and 29% I each ) 10 than to the various terpenoid synthases from the gymnosperm *Abies grandis* (monoterpene synthases with 49% S and 28% I (Bohlmann, J., Steele, C. L. and Croteau, R. (1997) *J. Biol. Chem.* **272**, 21784-21792); sesquiterpene synthases with 53% S and 29% I; diterpene synthases with 51% S and 28% I (Stofer Vogel, B., Wildung, M. R., Vogel, G. and Croteau, R. (1996) *J. Biol. Chem.* **271**, 23262- 15 20 23268). Even a phylogenetically distant diterpene cyclase from *Taxus brevifolia*, taxadiene synthase (Wildung, M. R. and Croteau, R. (1996) *J. Biol. Chem.* **271**, 9201-9204), resembles (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) at the amino acid level (50% S and 24% I) as closely as do the monoterpene synthases of the mint family. These sequence-based relationships may reflect a bifurcation in the evolution 25 of the monoterpene synthases from the higher terpenoid synthases that is as ancient as the separation between the angiosperms and gymnosperms.

#### Example 4

##### Characterization of (*E*)- $\beta$ -Farnesene Synthase (SEQ ID NO:2)

For determination of the pH optimum of (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2), the preparation was adjusted with 50 mM Mops (to a pH of 6.5, 6.75, 7.0, 30 7.25, 7.5, 8.0, or 8.5) before the assay. Kinetic constants for FDP, GDP, Mg<sup>++</sup> and Mn<sup>++</sup> were determined using a preparation of (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) that was partially purified by anion-exchange chromatography (on a Mono-Q column (Pharmacia) equilibrated with assay buffer and eluted with a linear KC1 gradient (0 to 500 mM) in assay buffer). The 210-230 mM fraction containing the 35

(*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) was used for kinetic evaluation of FDP and GDP as substrates (concentration range 0.31 to 20  $\mu$ M, with saturating  $Mg^{++}$ ). Due to the tenacious binding of divalent cations by the synthase, the partially purified enzyme (prepared in the presence of 10 mM EDTA) was dialyzed overnight against assay buffer containing 50 mM EDTA. The dialysate was buffer-exchanged by ultrafiltration (Amicon Centriprep 30, 450 fold dilution), then desalted (Bio-Rad Econo-Pak 10 DG) into assay buffer. Kinetic constants for  $Mg^{++}$  and  $Mn^{++}$  (assay range 1  $\mu$ M to 2 mM of the chloride salts) were then determined at 7.3  $\mu$ M [ $1\text{-}^3\text{H}$ ]FDP. Triplicate assays were conducted and control incubations (without enzyme) were included in all cases. A double reciprocal plot (Lineweaver, H. and Burk, D. (1934) *J. Am. Chem. Soc.* 56, 658-666) was generated for each averaged data set, and the equation of the best-fit line determined (Kaleidagraph ver. 3.08, Synergy Software).

The recombinant, partially purified (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) exhibited a broad pH optimum in the 6.75 to 7.25 range in Mopso buffer. This observation is in agreement with the studies of Salin et al. (Salin, F., Pauly, G., Charon, J. and Gleizes, M. (1995) *J. Plant Phys.* 146, 203-209) in which the purified (*E*)- $\beta$ -farnesene synthase from maritime pine was shown to possess a pH optimum in the 7.0 to 7.3 range. The  $K_m$  value for FDP with the recombinant synthase (SEQ ID NO:2) was calculated to be 0.6  $\mu$ M, a value typical of other sesquiterpene synthases of plant origin (Cane, D. E. (1990) *Chem. Rev.* 90, 1089-1103) but lower than the value of 5  $\mu$ M determined for the enzyme from maritime pine (Salin, F., Pauly, G., Charon, J. and Gleizes, M. (1995) *J. Plant Phys.* 146, 203-209). Substrate concentrations in excess of 10  $\mu$ M FDP evidenced slight inhibition of activity with the recombinant enzyme (SEQ ID NO:2). Although the relative velocity at saturating levels of GDP was only 3% of the velocity with FDP for the recombinant synthase (SEQ ID NO:2), the calculated  $K_m$  value for GDP (1.5  $\mu$ M) was only three-fold higher than that for FDP, suggesting that the binding of the  $C_{10}$  analog was reasonably efficient.

A  $K_m$  value of 150  $\mu$ M was determined for  $Mg^{++}$  ( $V_{rel} = 100$ ), and a  $K_m$  value of 7.0  $\mu$ M was determined for  $Mn^{++}$  ( $V_{rel} = 80$ ). No inhibition of activity was observed at  $Mg^{++}$  concentrations up to 10 mM; however, concentrations of  $Mn^{++}$  exceeding 20  $\mu$ M resulted in a sharp decline in reaction velocity to a plateau ( $V_{rel} = 20$ ) in the 0.25 to 2 mM range. Since the product distribution of the recombinant (*E*)- $\beta$ -farnesene synthase (SEQ ID NO:2) had been initially determined in the presence of

excess Mg<sup>++</sup>, the conversion of [1-<sup>3</sup>H]FDP was re-evaluated in the presence of Mn<sup>++</sup> alone at apparent saturation (20 μM). The olefin products were again analyzed by GC-MS and found in this case to consist of 98% (*E*)-β-farnesene and 2% (*Z*)-β-farnesene. No δ-cadinene, or other sesquiterpenes, were synthesized in this instance,  
5 indicating that a structural alteration in the binding of Mn<sup>++</sup> to the substrate and/or enzyme (relative to Mg<sup>++</sup>) improves the fidelity of the reaction.

In operational characteristics (pH optimum, kinetic constants) and physical features (size, pI), the recombinant (*E*)-β-farnesene synthase (SEQ ID NO:2) is a typical sesquiterpene synthase (Cane, D. E. (1990) *Chem. Rev.* **90**, 1089-1103;  
10 Fachinni, P. J. and Chappell, J. (1992) *Proc. Natl. Acad. Sci. USA* **89**, 11088-11092; Back, K. and Chappell, J. (1995) *J. Biol. Chem.* **270**, 7375-7381; Chen, X. Y., Chen, Y., Heinstein, P. and Davisson, V.J. (1996) *Arch. Biochem. Biophys.* **324**, 255-266), suggesting that the enzyme should be highly functional *in planta*. Given that this synthase (SEQ ID NO:2) will be targeted by default to the cytoplasm (Chappell, J.  
15 (1995) *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **46**, 521-547; Keegstra, K., Olsen, J. J. and Theg, S. M. (1989) *Ann. Rev. Plant Physiol. Plant Mol. Biol.* **40**, 471-501), where the substrate arises from the mevalonate pathway, it should be possible to engineer virtually any plant for the production of (*E*)-β-farnesene in order to exploit the kairomonal and pheromonal properties of this natural product.

20

### Example 5

#### Properties of (*E*)-β-Farnesene Synthase Proteins of the Present Invention

The (*E*)-β-farnesene synthase proteins of the present invention all require a divalent metal ion as a cofactor. Most (*E*)-β-farnesene synthase proteins of the present invention utilize either Mg<sup>++</sup> or Mn<sup>++</sup> as a cofactor. Nonetheless, (*E*)-β-farnesene synthase proteins of the present invention are inhibited at concentrations of Mn<sup>++</sup> in excess of about 5 mM.  
25

(*E*)-β-farnesene synthase proteins of the present invention have a pH optimum in the range of from about pH 5.5 to about pH 8.5, and a pI in the range of from about pH 4.5 to about pH 6.0. The Km(FPP) of (*E*)-β-farnesene synthase proteins of the present invention is less than about 10μM, while the Kcat(FPP) of (*E*)-β-farnesene synthase proteins of the present invention is less than about 5/sec. The (*E*)-β-farnesene synthase proteins of the present invention exist as either monomers or homodimers, with the monomer having a molecular weight of from about 55 kD (kiloDaltons) to about 65 kD.  
30

35

**Example 6****Hybridization of Peppermint (*E*)- $\beta$ -Farnesene Synthase cDNA (SEQ ID NO:1) to Other Nucleic Acid Sequences of the Present Invention**

The nucleic acid molecules of the present invention are capable of hybridizing  
5 to the nucleic acid sequence set forth in SEQ ID NO:1, or to the complementary  
sequence of the nucleic acid sequence set forth in SEQ ID NO:1, under the following  
stringent hybridization conditions: incubation in 5 X SSC at 65°C for 16 hours,  
followed by washing under the following conditions: two washes in 2 X SSC at 18°C  
10 to 25°C for twenty minutes per wash; preferably, two washes in 2 X SSC at 18°C to  
25°C for twenty minutes per wash, followed by one wash in 0.5 X SSC at 55°C for  
thirty minutes; most preferably, two washes in 2 X SSC at 18°C to 25°C for fifteen  
15 minutes per wash, followed by two washes in 0.2 X SSC at 65°C for twenty minutes  
per wash.

The ability of the nucleic acid molecules of the present invention to hybridize  
15 to the nucleic acid sequence set forth in SEQ ID NO:1, or to the complementary  
sequence of the nucleic acid sequence set forth in SEQ ID NO:1, can be determined  
utilizing the technique of hybridizing radiolabelled nucleic acid probes to nucleic acids  
immobilized on nitrocellulose filters or nylon membranes as set forth, for example, at  
pages 9.52 to 9.55 of Molecular Cloning, A Laboratory Manual (2nd edition), J.  
20 Sambrook, E.F. Fritsch and T. Maniatis eds, the cited pages of which are  
incorporated herein by reference.

In addition to the nucleic acid sequence set forth in SEQ ID NO:1, examples  
of representative nucleic acid sequences of the present invention that encode a  
peppermint (*E*)- $\beta$ -farnesene synthase protein and which hybridize to the  
25 complementary sequence of the nucleic acid sequence disclosed in SEQ ID NO:1 are  
set forth in SEQ ID NO:4; SEQ ID NO:6; SEQ ID NO:8; SEQ ID NO:10; SEQ ID  
NO:12; SEQ ID NO:14; SEQ ID NO:16 and SEQ ID NO:18. With the exception of  
the nucleic acid sequence set forth in SEQ ID NO:1, the foregoing representative  
nucleic acid sequences of the present invention were generated using a computer. By  
30 utilizing the degeneracy of the genetic code, each of the foregoing, representative  
nucleic acid sequences has a different sequence, but each encodes the protein set forth  
in SEQ ID NO:2. Thus, the identical (*E*)- $\beta$ -farnesene synthase protein sequence is set  
forth in SEQ ID NO:2, SEQ ID NO:5; SEQ ID NO:7; SEQ ID NO:9; SEQ ID  
NO:11; SEQ ID NO:13; SEQ ID NO:15; SEQ ID NO:17 and SEQ ID NO:19.

In addition to the protein sequence set forth in SEQ ID NO:2 examples of representative (*E*)- $\beta$ -farnesene synthase proteins of the present invention are set forth in SEQ ID NO:20; SEQ ID NO:21; SEQ ID NO:22; SEQ ID NO:23; SEQ ID NO:24; SEQ ID NO:25; SEQ ID NO:26, SEQ ID NO:27 and SEQ ID NO:28. With  
5 the exception of the amino acid sequence set forth in SEQ ID NO:2, the foregoing representative amino acid sequences of the present invention were generated using a computer by making conservative amino acid substitutions.

While the preferred embodiment of the invention has been illustrated and described, it will be appreciated that various changes can be made therein without  
10 departing from the spirit and scope of the invention.

The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

1. An isolated nucleic acid molecule encoding an (E)- $\beta$ -farnesene synthase protein.
2. An isolated nucleic acid molecule of Claim 1 encoding an angiosperm (E)- $\beta$ -farnesene synthase protein.
3. An isolated nucleic acid molecule of Claim 1 encoding a gymnosperm (E)- $\beta$ -farnesene synthase protein.
4. An isolated nucleic acid molecule of Claim 1 encoding an essential oil plant species (E)- $\beta$ -farnesene synthase protein.
5. An isolated nucleic acid molecule of Claim 1 encoding an (E)- $\beta$ -farnesene synthase protein from the genus *Mentha*.
6. An isolated nucleic acid molecule of Claim 5 encoding an (E)- $\beta$ -farnesene synthase protein from *Mentha piperita*.
7. An isolated nucleic acid molecule of Claim 6 consisting of the nucleic acid sequence set forth in SEQ ID NO:1.
8. An isolated nucleic acid molecule of Claim 1 encoding an (E)- $\beta$ -farnesene synthase protein having the amino acid sequence set forth in SEQ ID NO:2.
9. An isolated (E)- $\beta$ -farnesene synthase protein, provided that said isolated (E)- $\beta$ -farnesene synthase protein is not native to Maritime pine.
10. A gymnosperm (E)- $\beta$ -farnesene synthase protein of Claim 9.
11. An angiosperm (E)- $\beta$ -farnesene synthase protein of Claim 9.
12. An essential oil plant (E)- $\beta$ -farnesene synthase protein of Claim 9.
13. A *Mentha* (E)- $\beta$ -farnesene synthase protein of Claim 9.
14. A *Mentha piperita* (E)- $\beta$ -farnesene synthase protein of Claim 13.

15. An (E)- $\beta$ -farnesene synthase protein of Claim 13, said protein consisting of the amino acid sequence set forth in SEQ ID NO:2.

16. A replicable expression vector comprising a nucleic acid sequence encoding an (E)- $\beta$ -farnesene synthase protein.

17. A replicable expression vector of Claim 16 comprising a nucleic acid sequence encoding an angiosperm (E)- $\beta$ -farnesene synthase protein.

18. A replicable expression vector of Claim 16 comprising a nucleic acid sequence encoding a gymnosperm (E)- $\beta$ -farnesene synthase protein.

19. A replicable expression vector of Claim 16 comprising a nucleic acid sequence encoding an essential oil plant (E)- $\beta$ -farnesene synthase protein.

20. A replicable expression vector of Claim 16 comprising a nucleic acid sequence encoding a *Mentha* (E)- $\beta$ -farnesene synthase protein.

21. A replicable expression vector of Claim 16 comprising a nucleic acid sequence encoding a *Mentha piperita* (E)- $\beta$ -farnesene synthase protein.

22. A replicable expression vector of Claim 16 comprising a nucleic acid sequence consisting of the nucleic acid sequence set forth in SEQ ID NO:1.

23. A host cell comprising a vector of Claim 16.

24. A host cell comprising a vector of Claim 17.

25. A host cell comprising a vector of Claim 18.

26. A host cell comprising a vector of Claim 19.

27. A host cell comprising a vector of Claim 20.

28. A host cell comprising a vector of Claim 21.

29. A host cell comprising a vector of Claim 22.

30. A host cell of Claim 23, said host cell being a plant cell.

31. An isolated nucleic acid molecule that is capable of hybridizing to the nucleic acid molecule set forth in SEQ ID NO:1, or to the complementary sequence of the nucleic acid molecule set forth in SEQ ID NO:1, under stringent hybridization conditions.

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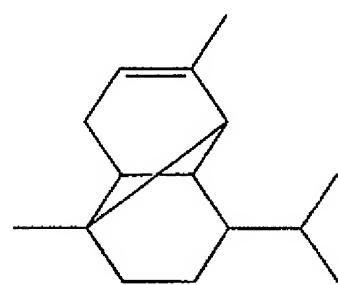
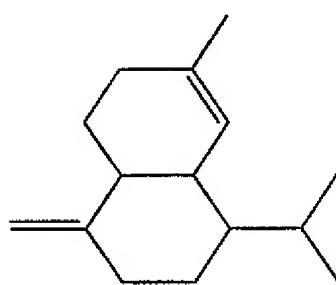
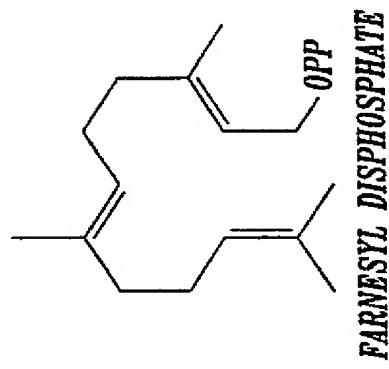
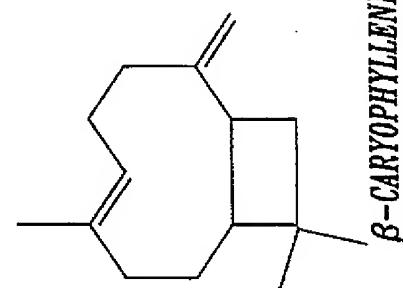
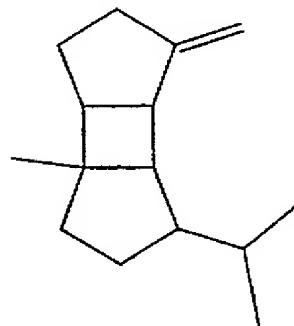
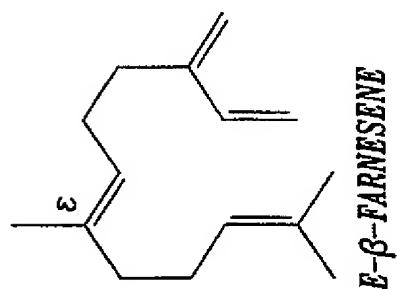
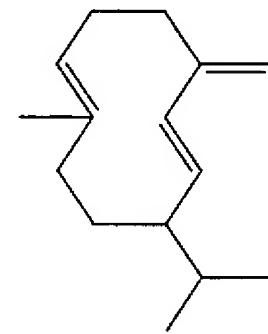
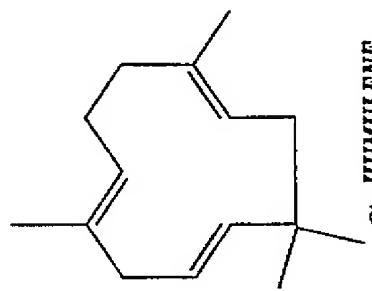
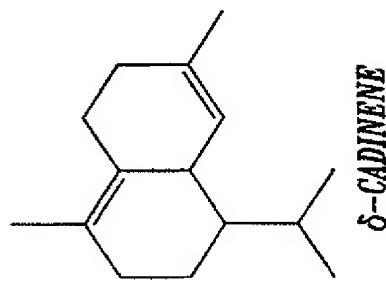


Fig. 1.

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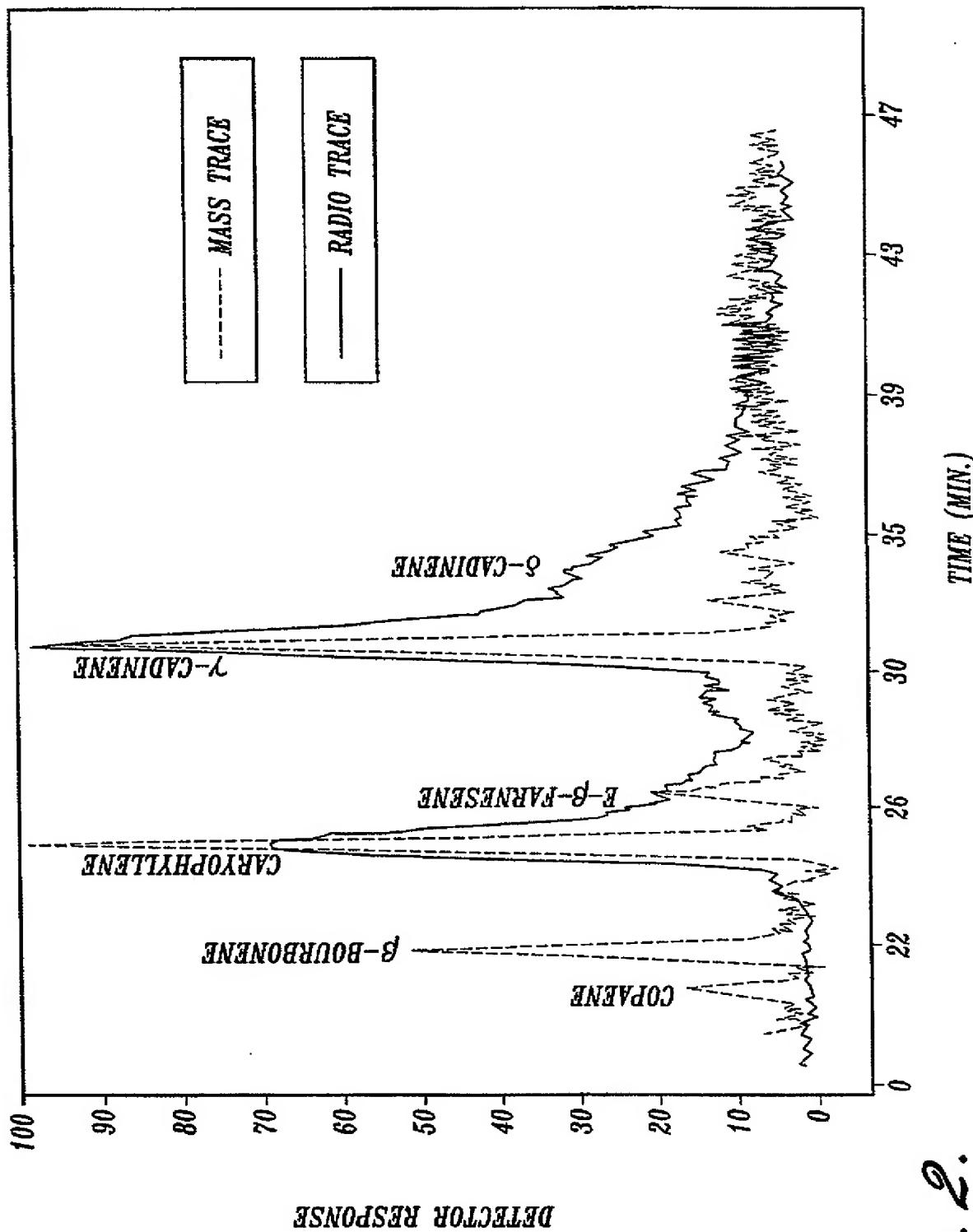


Fig. 2.

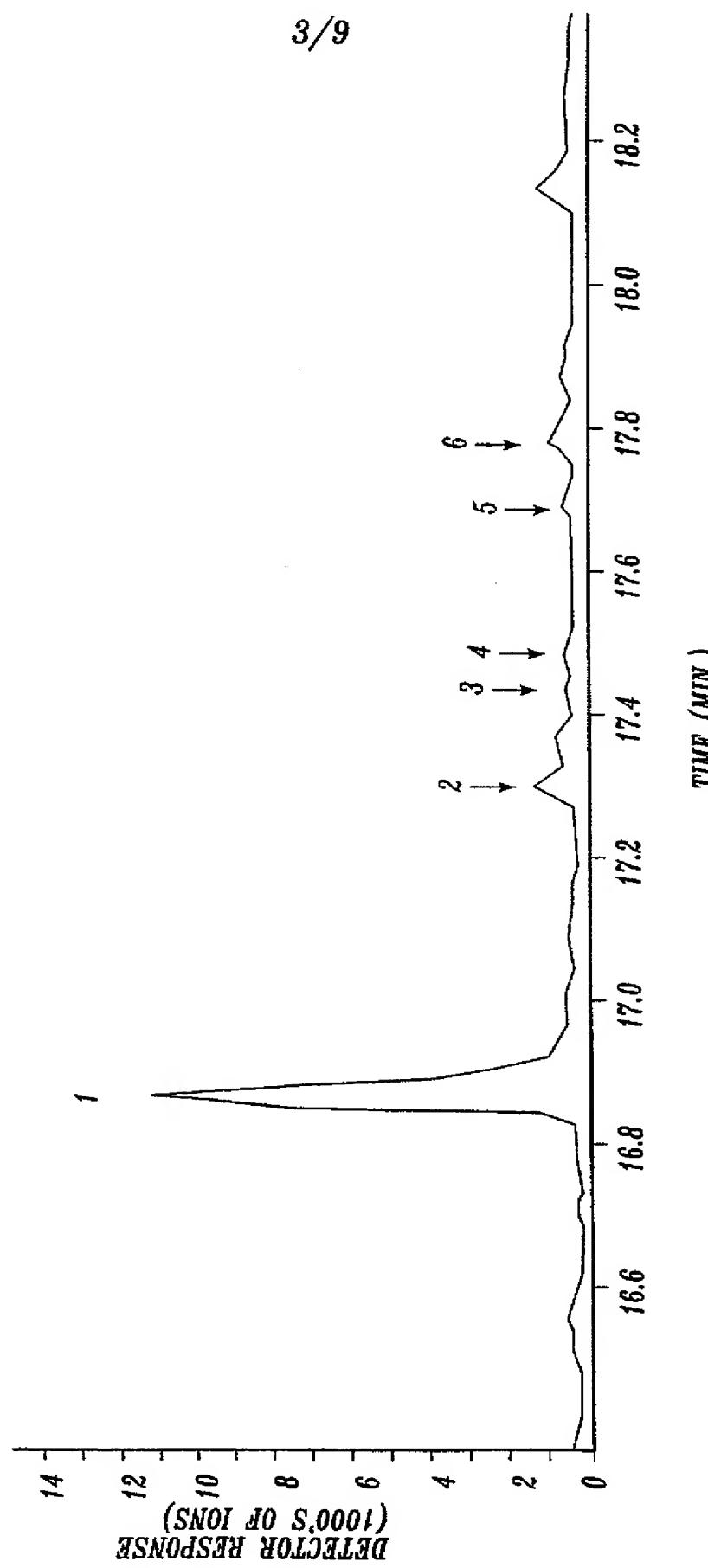


Fig. 9.d.

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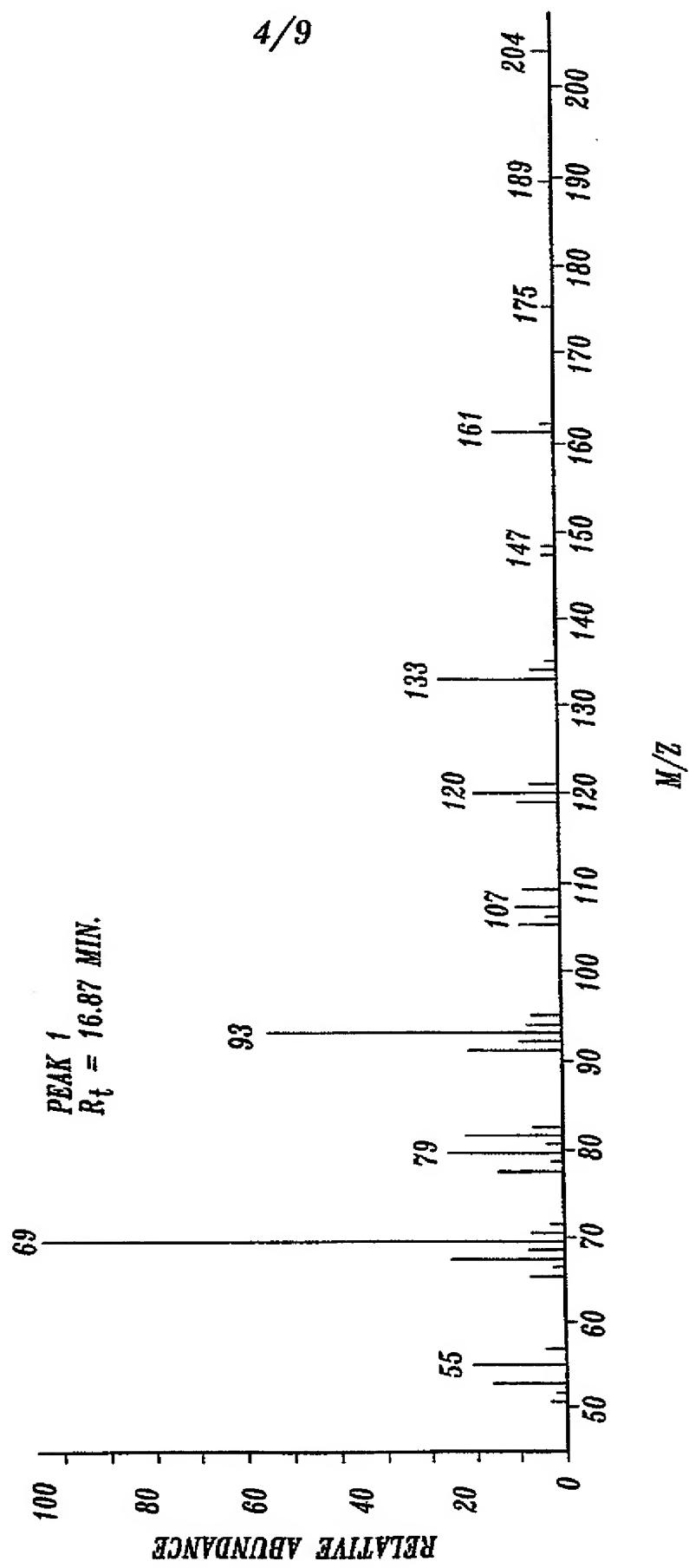
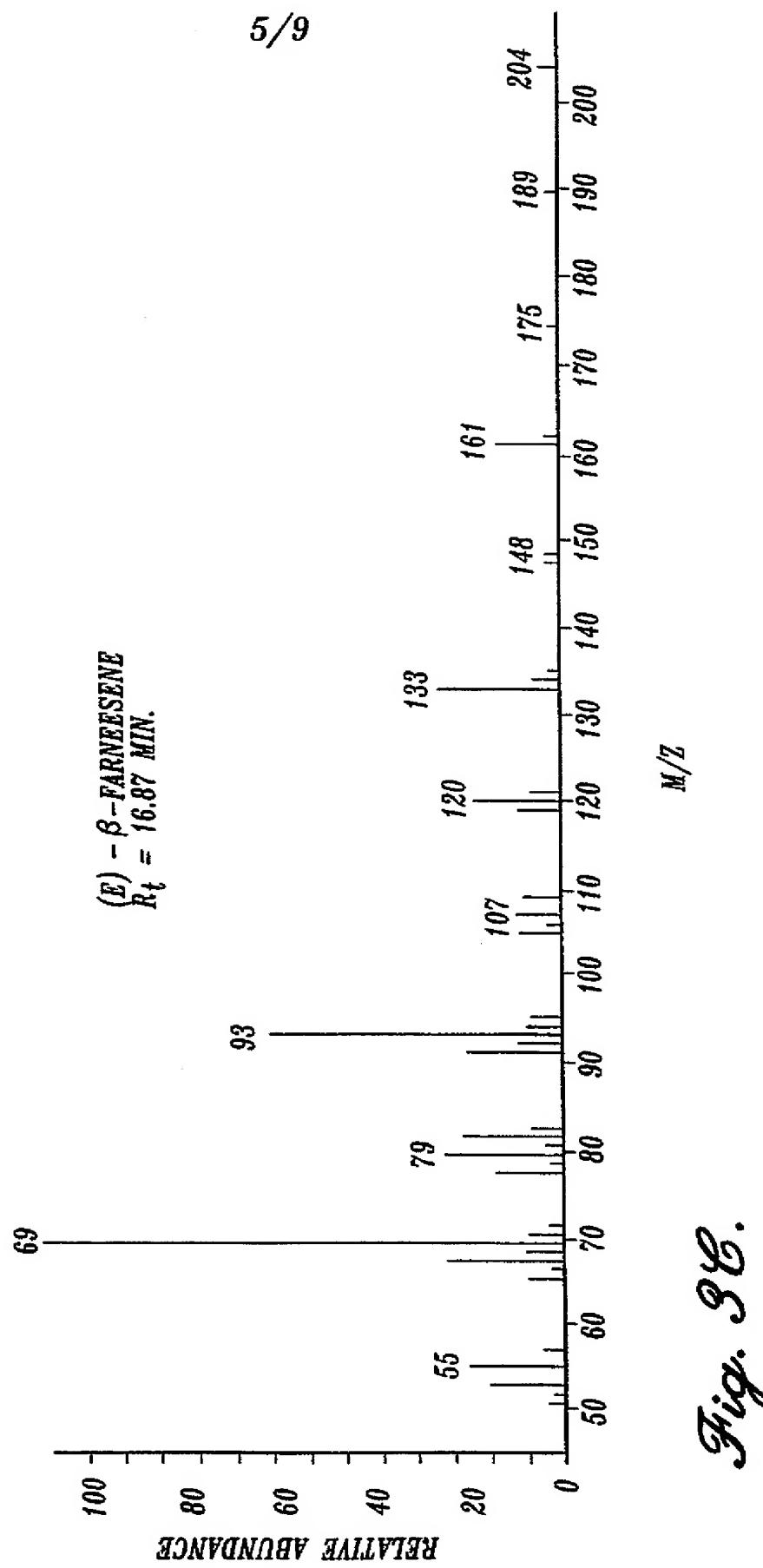


Fig. 3B.

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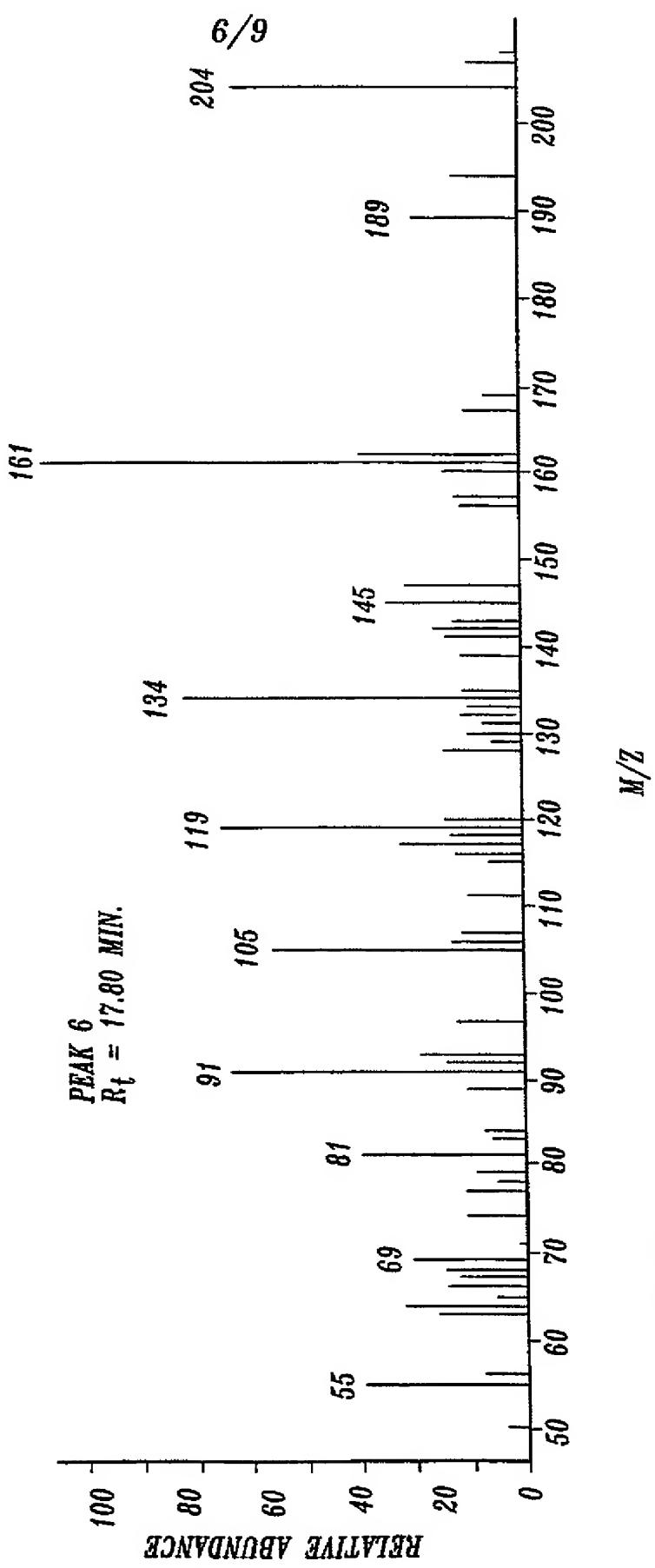


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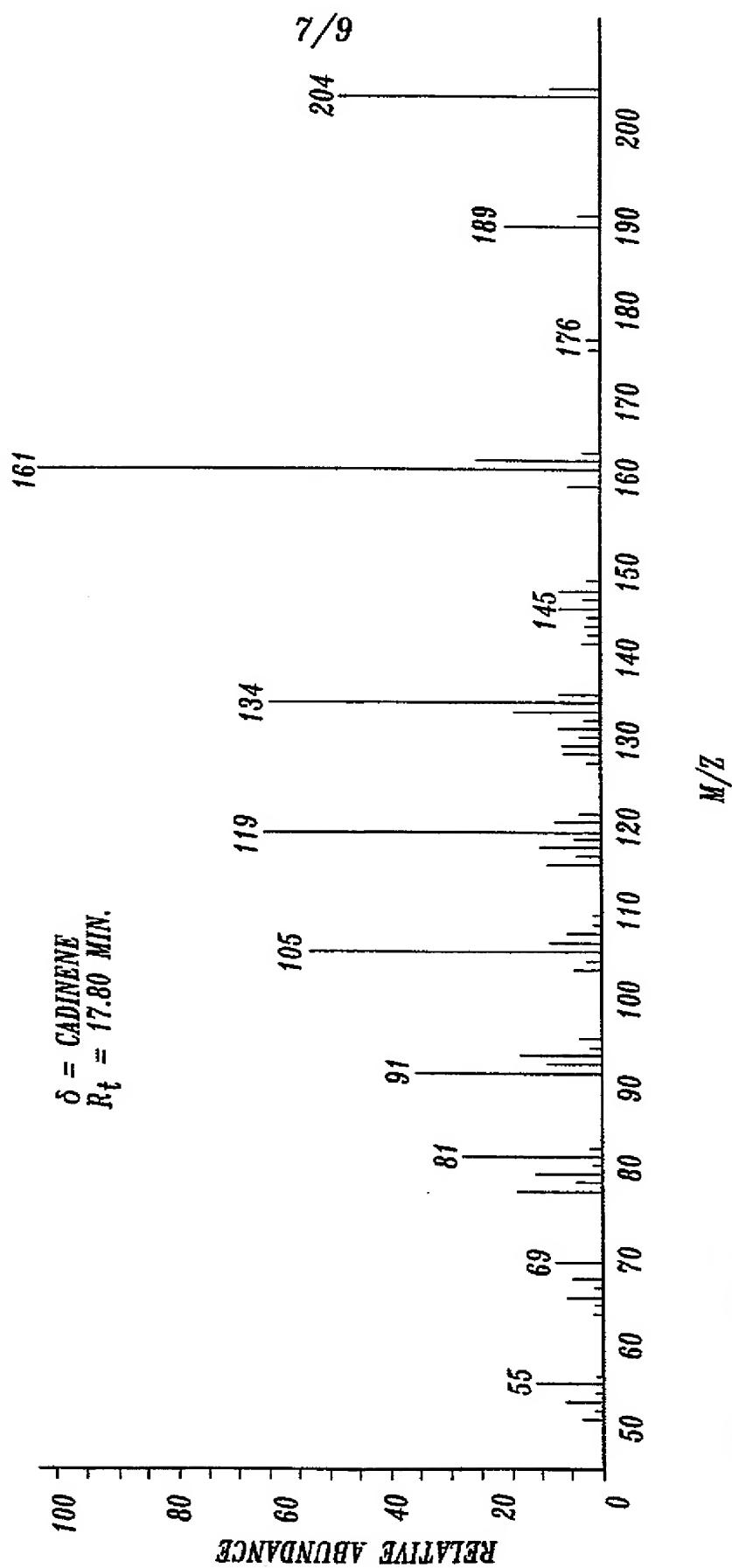
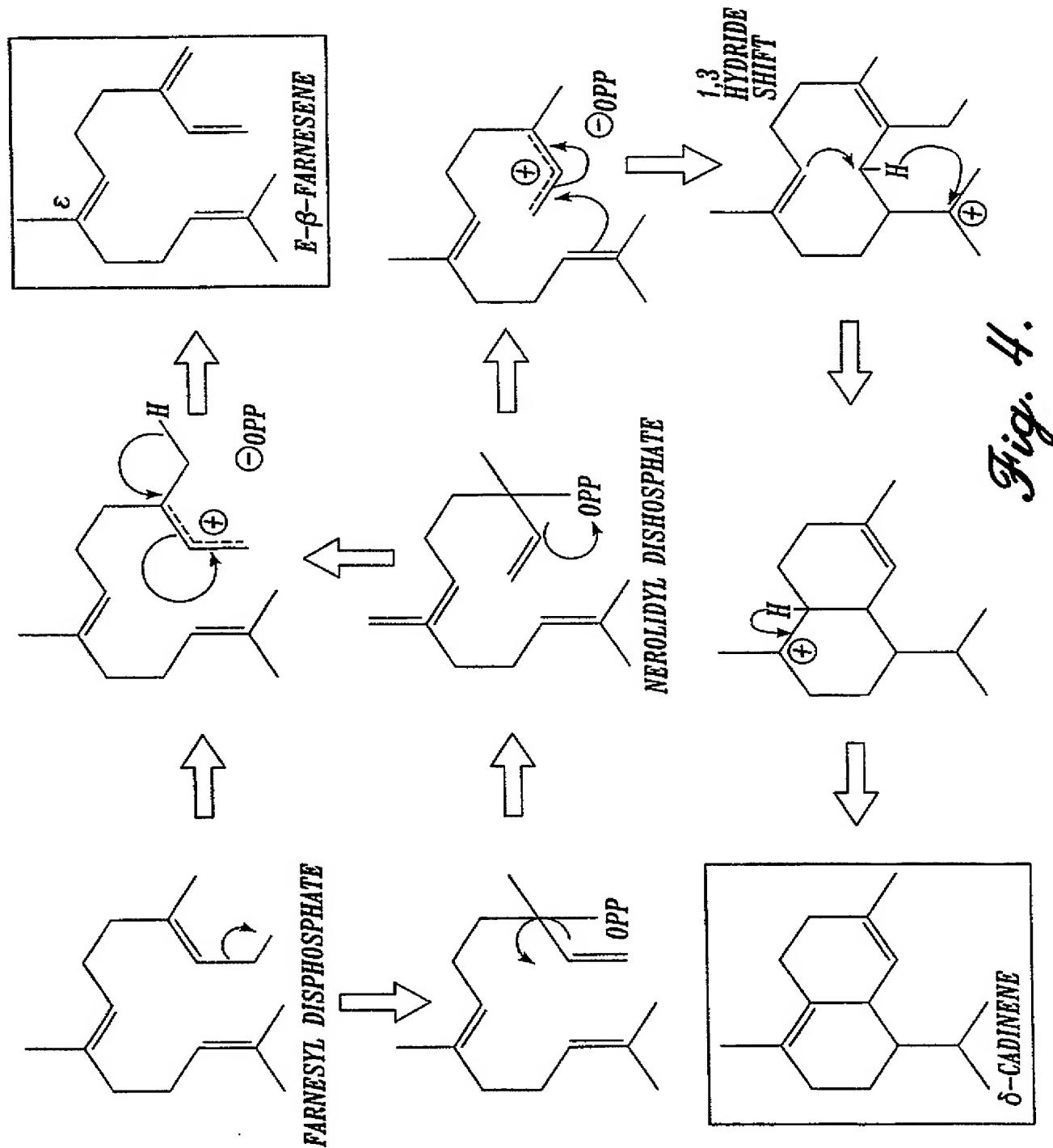
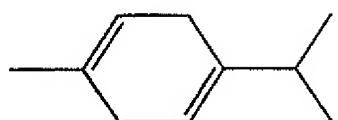


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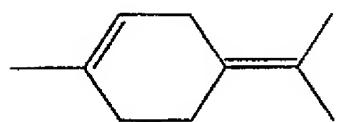
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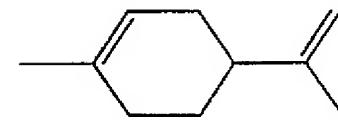
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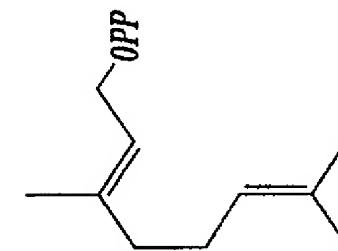
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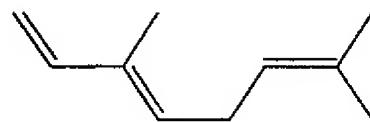
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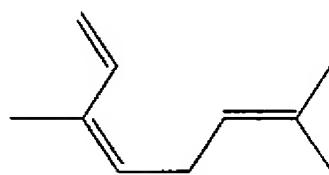
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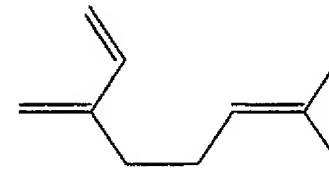
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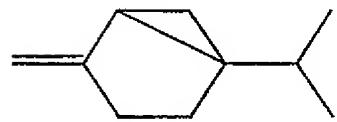
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Z-OCIMENTE



MYRCENE



SAHINENE

Fig. 5.

## SEQUENCE LISTING

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Wildung, Mark R  
Crock, John E

<120> Isolation and Bacterial Expression of a Sesquiterpene Synthase cDNA Clone from Peppermint (*Mentha x piperita*, L.) that Produces the Aphid Alarm Pheromone (E)-beta-Farnesene

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<170> PatentIn Ver. 2.0

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<211> 1959  
<212> DNA  
<213> *Mentha piperita*

<220>

<221> CDS

<222> (71)..(1720)

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Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu  
1 5 10

gta agg cca cct atg acg aag cat gcg cca agc atg tgg act gat acc 157  
Val Arg Pro Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr  
15 20 25

ttt tct aac ttt tct ctt gac gat aag gaa caa caa aag tgc tca gaa 205  
Phe Ser Asn Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu  
30 35 40 45

acc atc gaa gca ctt aag caa gaa gca aga ggc atg ctt atg gct gca 253

Thr Ile Glu Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala  
 50 55 60

acc act cct ctc caa caa atg aca cta atc gac act ctc gag cgt ttg 301  
 Thr Thr Pro Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu  
 65 70 75

gga ttg tct ttc cat ttt gag acg gag atc gaa tac aaa atc gaa cta 349  
 Gly Leu Ser Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu  
 80 85 90

atc aac gct gca gaa gac gac ggc ttt gat ttg ttc gct act gct ctt 397  
 Ile Asn Ala Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu  
 95 100 105

cgt ttc cgt ttg ctc aga caa cat caa cgc cac gtt tct tgt gat gtt 445  
 Arg Phe Arg Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val  
 110 115 120 125

ttc gac aag ttc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc 493  
 Phe Asp Lys Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser  
 130 135 140

aat aat gtt gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg 541  
 Asn Asn Val Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly  
 145 150 155

ttt cgc gaa gaa aga ata tta caa gag gct gta aat ttt acg agg cat 589  
 Phe Arg Glu Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His  
 160 165 170

cac ttg gaa gga gca gag tta gat cag tct cca tta ttg att aga gag 637  
 His Leu Glu Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu  
 175 180 185

aaa gtg aag cga gct ttg gag cac cct ctt cat agg gat ttc ccc att 685  
 Lys Val Lys Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile  
 190 195 200 205

gtc tat gca cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga 733  
 Val Tyr Ala Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg  
 210 215 220

gat gaa tta ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag 781  
 Asp Glu Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln  
 225 230 235

aat ttg tat aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca 829

Asn Leu Tyr Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr			
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tgg aat ctg aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag			877
Trp Asn Leu Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu			
255	260	265	
gct tat gtt tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat			925
Ala Tyr Val Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr			
270	275	280	285
gtt cga atg gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac			973
Val Arg Met Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp			
290	295	300	
gat aca tat gat aat tat gct aca ctc aat gaa gct caa ctt ttt act			1021
Asp Thr Tyr Asp Asn Tyr Ala Thr Ieu Asn Glu Ala Gln Leu Phe Thr			
305	310	315	
caa gtc tta gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa			1069
Gln Val Leu Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu			
320	325	330	
tac atg aaa atc gtt tat cga ttt att ttg agt ata tat gaa aat tat			1117
Tyr Met Lys Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr			
335	340	345	
gaa cgt gat gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt			1165
Glu Arg Asp Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe			
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aag gaa acc gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag			1213
Lys Glu Thr Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys			
370	375	380	
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Trp Val Met Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn			
385	390	395	
tca gag aaa acc agc tgc att tat acc atg ttt gct tct atc atc cca			1309
Ser Glu Lys Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro			
400	405	410	
ggc ttg aaa tct gtt acc caa gaa acc att gat tgg atc aag agt gaa			1357
Gly Leu Lys Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu			
415	420	425	
ccc acg ctc gca aca tcg acc gct atg atc ggt cgg tat tgg aat gac			1405

Pro Thr Leu Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp  
 430 435 440 445

acc agc tct cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg 1453  
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 450 455 460

ttg gat ttc cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca 1501  
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 465 470 475

tct aag ttt gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag 1549  
 Ser Lys Phe Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys  
 480 485 490

gaa ttc ata gcc aca act aat tat aat gtg ggt aga gaa att gcc atc 1597  
 Glu Phe Ile Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile  
 495 500 505

aca ttc ctc aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act 1645  
 Thr Phe Leu Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr  
 510 515 520 525

gac gga gac gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt 1693  
 Asp Gly Asp Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val  
 530 535 540

gct ctc ttt gtt gat gcc ata gtc ttt tgatttgcat aatcaaagac 1740  
 Ala Leu Phe Val Asp Ala Ile Val Phe  
 545 550

cctataatta taatttatatg tgtttaagaa actaataagc ttgctttatg tatagttgac 1800  
 aattgaataa taatgtatata attagtagag ttaagaagtt ataaaagaata aagaggagct 1860  
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 ttggaagaga ccaatatata caaaaaaaaaaaaaaaa 1959

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<212> PRT  
<213> Mentha piperita

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35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
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Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
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Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
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Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
305 310 315 320  
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
325 330 335  
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
340 345 350  
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365  
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380  
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385 390 395 400  
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415  
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430  
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445  
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460  
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480  
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495  
Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510  
Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

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Val Asp Ala Ile Val Phe  
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amino acid motif

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<221> DOMAIN  
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<223> Conserved domain that may coordinate binding of  
divalent metal ion

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sequence encoding peppermint E-beta-farnesene  
synthase protein

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peppermint E-beta-farnesene synthase protein

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cct atg acg aag cat gcg cca agc atg tgg act gat acc ttt tct aac 96  
 Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
 20 25 30

ttt tct ctt gac gat aag gaa caa caa aag tgc tca gaa acc atc gaa 144  
 Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35 40 45

gca ctt aag caa gaa gca aga ggc atg ctt atg gct gca acc act cct 192  
 Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50 55 60

ctc caa caa atg aca cta atc gac act ctc gag cgt ttg gga ttg tct 240  
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 65 70 75 80

tcc cat ttt gag acg gag atc gaa tac aaa atc gaa cta atc aac gct 288  
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gca gaa gac gac ggc ttt gat ttg ttc gct act gct ctt cgt ttc cgt 336  
 Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

ttg ctc aga caa cat caa cgc cac gtt tct tgt gat gtt ttc gac aag 384  
 Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

tcc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc aat aat gtt 432  
 Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg ttt cgc gaa 480  
 Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa 528  
 Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag 576  
 Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca 624  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta 672  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat 720  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg 768  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt 816  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg 864  
 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat 912  
 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta 960  
 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa 1008  
 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat 1056  
 Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc 1104  
 Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg 1152  
 Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa 1200  
 Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa 1248  
 Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
         405                       410                       415

tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc 1296  
 Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
         420                       425                       430

gca aca tcg acc gct atg atc ggt cggt tat tgg aat gac acc agc tct 1344  
 Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
         435                       440                       445

cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392  
 Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
         450                       455                       460

cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt 1440  
 His Met Lys Glu Tyr Gly Leu Thr Lys Glu Ala Ala Ser Lys Phe  
         465                       470                       475                       480

gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata 1488  
 Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
         485                       490                       495

gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc 1536  
 Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
         500                       505                       510

aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
         515                       520                       525

gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
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gtt gat gcc ata gtc ttt  
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Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
 405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
 420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
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Val Asp Ala Ile Val Phe  
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<212> DNA  
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<220>  
<223> Description of Artificial Sequence: nucleic acid  
sequence encoding E-beta-farnesene synthase  
protein

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<221> CDS  
<222> (1)..(1650)  
<223> Computer-generated nucleic acid sequence encoding  
peppermint E-beta-farnesene synthase protein

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cct atg tcg aag cat gcg cca agc atg tgg act gat acc ttt tct aac      96  
Pro Met Ser Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20                    25                    30

ttt tct ctt gac gat aag gaa caa caa aag tgc tca gaa acc atc gaa      144  
Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35                    40                    45

gca ctt aag caa gaa gca aga ggc atg ctt atg gct gca acc acc cct      192  
Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50                    55                    60

ctc caa caa atg aca cta atc gac act ctc gag cgt ttg gga ttg tct      240  
Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65                    70                    75                    80

ttc cat ttt gag acg gag atc gaa tac aaa atc gaa cta atc aac gct      288

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85 90 95

gca gaa gac gac ggc ttt gat ttg ttc gct act gct ctt cgt ttc cgt 336  
 Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

ttg ctc aga caa cat caa cgc cac gtt tct tgt gat gtt ttc gac aag 384  
 Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

ttc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc aat aat gtt 432  
 Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg ttt cgc gaa 480  
 Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa 528  
 Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag 576  
 Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca 624  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta 672  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat 720  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg 768  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt 816  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg 864

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met			
275	280	285	
gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat			912
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr			
290	295	300	
gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta			960
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu			
305	310	315	320
gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa			1008
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys			
325	330	335	
atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat			1056
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp			
340	345	350	
gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc			1104
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr			
355	360	365	
gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg			1152
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met			
370	375	380	
gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa			1200
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys			
385	390	395	400
acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa			1248
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys			
405	410	415	
tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc			1296
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu			
420	425	430	
gca aca tcg acc gct atg atc ggt cgg tat tgg aat gac acc agc tct			1344
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser			
435	440	445	
cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc			1392
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe			
450	455	460	
cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt			1440

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465                   470                   475                   480  
 gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata   1488  
 Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485                   490                   495  
 gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc   1536  
 Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500                   505                   510  
 aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac   1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515                   520                   525  
 gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt   1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530                   535                   540  
 gtt gat gcc ata gtc ttt   1650  
 Val Asp Ala Ile Val Phe  
 545                   550  
  
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 Pro Met Ser Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
 20                   25                       30  
  
 Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35                   40                       45  
  
 Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50                   55                       60  
  
 Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65                   70                       75                       80  
  
 Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85                   90                       95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

&lt;210&gt; 8

&lt;211&gt; 1650

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: nucleic acid  
sequence encoding peppermint E-beta-farnesene

<220>  
 <221> CDS  
 <222> (1)..(1650)  
 <223> Computer-generated nucleic acid sequence encoding  
 peppermint E-beta-farnesene synthase protein

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atg	gct	aca	aac	ggc	gtc	gta	att	agt	tgc	tta	agg	gaa	gta	agg	cca	48
Met	Ala	Thr	Asn	Gly	Val	Val	Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro	
1	5									10					15	
cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20	25									25					30	
ttt	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Leu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	
35	40									40					45	
gca	ctt	aag	caa	gaa	gca	aga	ggc	atg	ctt	atg	gct	gca	acc	act	cct	192
Ala	Leu	Lys	Gln	Glu	Ala	Arg	Gly	Met	Leu	Met	Ala	Ala	Thr	Thr	Pro	
50	55									55					60	
ctc	caa	caa	atg	aca	cta	atc	gac	act	ctc	gag	cgt	ttg	gga	ttg	tct	240
Leu	Gln	Gln	Met	Thr	Leu	Ile	Asp	Thr	Leu	Glu	Arg	Leu	Gly	Leu	Ser	
65	70									75					80	
ttc	cat	ttt	gag	acg	gag	atc	gaa	tac	aaa	atc	gaa	cta	atc	aac	gct	288
Phe	His	Phe	Glu	Thr	Glu	Ile	Glu	Tyr	Lys	Ile	Glu	Leu	Ile	Asn	Ala	
85	90									90					95	
gca	gaa	gac	gac	ggc	ttt	gat	ttg	ttc	gct	act	gct	ctt	cgt	ttc	cgt	336
Ala	Glu	Asp	Asp	Gly	Phe	Asp	Leu	Phe	Ala	Thr	Ala	Leu	Arg	Phe	Arg	
100	105									105					110	
ttg	ctc	aga	caa	cat	caa	cgc	cac	gtt	tct	tgt	gat	gtt	ttc	gac	aag	384
Leu	Leu	Arg	Gln	His	Gln	Arg	His	Val	Ser	Cys	Asp	Val	Phe	Asp	Lys	
115	120									120					125	
ttc	atc	gac	aaa	gat	ggc	aag	ttc	gaa	gaa	tcc	ctt	agc	aat	aat	gtt	432
Phe	Ile	Asp	Lys	Asp	Gly	Lys	Phe	Glu	Glu	Ser	Leu	Ser	Asn	Asn	Val	
130	135									135					140	
gaa	ggc	cta	tta	agc	ttg	tat	gaa	gca	gct	cat	gtt	ggg	ttt	cgc	gaa	480
Glu	Gly	Leu	Leu	Ser	Leu	Tyr	Glu	Ala	Ala	His	Val	Gly	Phe	Arg	Glu	
145	150									150					160	

gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa 528  
 Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
     165                       170                       175  
  
 gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag 576  
 Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
     180                       185                       190  
  
 cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca 624  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
     195                       200                       205  
  
 cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta 672  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
     210                       215                       220  
  
 ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat 720  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
     225                       230                       235                       240  
  
 aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg 768  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
     245                       250                       255  
  
 aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt 816  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
     260                       265                       270  
  
 tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg 864  
 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
     275                       280                       285  
  
 gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat 912  
 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
     290                       295                       300  
  
 gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta 960  
 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
     305                       310                       315                       320  
  
 gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa 1008  
 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
     325                       330                       335  
  
 atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat 1056  
 Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
     340                       345                       350

gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc 1104  
 Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
                   355                  360                  365  
  
 gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg 1152  
 Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
                   370                  375                  380  
  
 gaa agg cag cta ccg tca ttccaa gac tac gta aag aat tca gag aaa 1200  
 Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
                   385                  390                  395                  400  
  
 acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa 1248  
 Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
                   405                  410                  415  
  
 tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc 1296  
 Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
                   420                  425                  430  
  
 gca aca tcg acc gct atg atc ggt cggt tat tgg aat gac acc agc tct 1344  
 Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
                   435                  440                  445  
  
 cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392  
 Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
                   450                  455                  460  
  
 cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt 1440  
 His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
                   465                  470                  475                  480  
  
 gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata 1488  
 Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
                   485                  490                  495  
  
 gcc aca act caa tat aat gtg ggt aga gaa att gcc atc aca ttc ctc 1536  
 Ala Thr Thr Gln Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
                   500                  505                  510  
  
 aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
                   515                  520                  525  
  
 gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
                   530                  535                  540

gtt gat gcc ata gtc ttt 1650  
 Val Asp Ala Ile Val Phe  
 545 550

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Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala

195	200	205
Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu		
210	215	220
Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr		
225	230	235
Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu		
245	250	255
Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val		
260	265	270
Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met		
275	280	285
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr		
290	295	300
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu		
305	310	315
320		
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys		
325	330	335
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp		
340	345	350
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr		
355	360	365
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met		
370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
400		
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		
435	440	445
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe		

450	455	460
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe		
465	470	475
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile		
485	490	495
Ala Thr Thr Gln Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu		
500	505	510
Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp		
515	520	525
Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe		
530	535	540
Val Asp Ala Ile Val Phe		
545	550	

&lt;210&gt; 10

&lt;211&gt; 1650

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: nucleic acid  
sequence encoding E-beta-farnesene synthase  
protein

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1)..(1650)

<223> Computer-generated nucleic acid sequence encoding  
peppermint E-beta-farnesene synthase protein

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atg	gtc	aca	aac	ggc	gtc	gta	att	agt	tgc	tta	agg	gaa	gta	agg	cca	48
Met		Ala		Gly		Val		Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro
1		5						10						15		

cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20		25												30		

tcc	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Leu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	

35	40	45	
gca ctt aag caa gaa gca aga ggc atg ctt atg gct gca acc act cct Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro			192
50	55	60	
ctc caa caa atg aca cta atc gac act ctc gag cgt ttg gga ttg tct Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser			240
65	70	75	80
ttc cat ttt gag acg gag atc gaa tac aaa atc gaa cta atc aac gct Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala			288
85	90	95	
gca gaa gac gac ggc ttt gat ttg ttc gct act gct ctt cgt ttc cgt Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg			336
100	105	110	
ttg ctc aga caa cat caa cgc cac gtt tcg tgt gat gtt ttc gac aag Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys			384
115	120	125	
ttc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc aat aat gtt Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val			432
130	135	140	
gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg ttt cgc gaa Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu			480
145	150	155	160
gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu			528
165	170	175	
gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys			576
180	185	190	
cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala			624
195	200	205	
cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu			672
210	215	220	
ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr			720

225	230	235	240	
aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu				768
245	250	255		
aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val				816
260	265	270		
tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met				864
275	280	285		
gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr				912
290	295	300		
gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu				960
305	310	315	320	
gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys				1008
325	330	335		
atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp				1056
340	345	350		
gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr				1104
355	360	365		
gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met				1152
370	375	380		
gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys				1200
385	390	395	400	
acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys				1248
405	410	415		
tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu				1296

420

425

430

gca aca tcg acc gct atg atc ggt cgg tat tgg aat gac acc agc tct 1344  
 Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392  
 Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt 1440  
 His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465 470 475 480

gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata 1488  
 Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc 1536  
 Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530 535 540

gtt gat gcc ata gtc ttt 1650  
 Val Asp Ala Ile Val Phe  
 545 550

<210> 11  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<400> 11  
Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu

35	40	45
Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro		
50	55	60
Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser		
65	70	75
Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala		
85	90	95
Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg		
100	105	110
Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys		
115	120	125
Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val		
130	135	140
Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu		
145	150	155
160		
Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu		
165	170	175
Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys		
180	185	190
Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala		
195	200	205
Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu		
210	215	220
Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr		
225	230	235
240		
Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu		
245	250	255
Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val		
260	265	270
Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met		
275	280	285
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr		

290	295	300
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu		
305	310	315
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys		
325	330	335
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp		
340	345	350
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr		
355	360	365
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met		
370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		
435	440	445
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe		
450	455	460
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe		
465	470	475
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile		
485	490	495
Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu		
500	505	510
Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp		
515	520	525
Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe		
530	535	540
Val Asp Ala Ile Val Phe		

545

550

&lt;210&gt; 12

&lt;211&gt; 1650

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: nucleic acid sequence encoding E-beta-farnesene synthase

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1)..(1650)

&lt;223&gt; Computer-generated nucleic acid sequence encoding peppermint E-beta-farnesene synthase protein

&lt;400&gt; 12

atg	gct	ggg	aac	ggc	gtc	gta	att	agt	tgc	tta	agg	gaa	gta	agg	cca	48
Met	Ala	Gly	Asn	Gly	Val	Val	Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro	
1					5				10					15		

cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20						25				30						

ttt	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Ieu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	
35					40				45							

gca	ctt	aag	caa	gaa	gca	aga	ggc	atg	ctt	atg	gct	gca	acc	act	cct	192
Ala	Leu	Lys	Gln	Glu	Ala	Arg	Gly	Met	Leu	Met	Ala	Ala	Thr	Thr	Pro	
50					55				60							

ctc	caa	caa	atg	aca	cta	atc	gac	act	ctc	gag	cgt	ttg	gga	ttg	tct	240
Leu	Gln	Gln	Met	Thr	Ieu	Ile	Asp	Thr	Ieu	Glu	Arg	Leu	Gly	Ieu	Ser	
65					70				75			80				

ttc	cat	ttt	gag	acg	gag	atc	gaa	tac	aaa	atc	gaa	cta	atc	aac	gct	288
Phe	His	Phe	Glu	Thr	Glu	Ile	Glu	Tyr	Lys	Ile	Glu	Ieu	Ile	Asn	Ala	
85					90				95							

gca	gaa	gac	gac	ggc	ttt	gat	ttg	ttc	gct	act	gct	ctt	cgt	ttc	cgt	336
Ala	Glu	Asp	Asp	Gly	Phe	Asp	Ieu	Phe	Ala	Thr	Ala	Leu	Arg	Phe	Arg	
100					105				110							

ttg	ctc	aga	caa	cat	caa	cgc	cac	gtt	tct	tgt	gat	gtt	ttc	gac	aag	384
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys			
115	120	125	
 ttc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc aat aat gtt			432
Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val			
130	135	140	
 gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg ttt cgc gaa			480
Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu			
145	150	155	160
 gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa			528
Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu			
165	170	175	
 gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag			576
Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys			
180	185	190	
 cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca			624
Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala			
195	200	205	
 cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta			672
Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu			
210	215	220	
 ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat			720
Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr			
225	230	235	240
 aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg			768
Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu			
245	250	255	
 aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt			816
Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val			
260	265	270	
 tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg			864
Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met			
275	280	285	
 gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat			912
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr			
290	295	300	
 gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta			960

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu				
305	310	315	320	
gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa				1008
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys				
325	330	335		
atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat				1056
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp				
340	345	350		
gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc				1104
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr				
355	360	365		
gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg				1152
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Lys Trp Val Met				
370	375	380		
gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa				1200
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys				
385	390	395	400	
acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa				1248
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys				
405	410	415		
tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc				1296
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu				
420	425	430		
gca aca tcg acc gct atg atc ggt cgg tat tgg aat gac acc agc tct				1344
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser				
435	440	445		
cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc				1392
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe				
450	455	460		
cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt				1440
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe				
465	470	475	480	
gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata				1498
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile				
485	490	495		
gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc				1536

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530 535 540

gtt gat gcc ata gtc ttt 1650  
 Val Asp Ala Ile Val Phe  
 545 550

<210> 13  
 <211> 550  
 <212> PRT  
 <213> Artificial Sequence

<400> 13  
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 1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
 20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160  
 Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175  
 Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270  
 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285  
 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300  
 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320  
 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335  
 Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350  
 Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365  
 Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380  
 Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

<210> 14

<211> 1650

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: nucleic acid  
sequence encoding E-beta-farnesene synthase  
protein

<220>

<221> CDS

<222> (1)..(1650)

<223> Computer-generated nucleic acid sequence encoding  
peppermint E-beta-farnesene synthase protein

<400> 14

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Met		Ala	Thr	Asn	Gly	Val	Val	Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro
1		5						10							15	

cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20		25							30							

ttt	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Leu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	
35		40							45							

gca	ctt	aag	caa	gaa	gca	aga	ggc	atg	ctt	atg	gct	gca	acc	act	cct	192
Ala	Leu	Lys	Gln	Glu	Ala	Arg	Gly	Met	Leu	Met	Ala	Ala	Thr	Thr	Pro	
50		55							60							

ctc	caa	caa	atg	aca	cta	atc	gac	act	ctc	gag	cgt	ttg	gga	ttg	tct	240
Leu	Gln	Gln	Met	Thr	Ile	Asp	Thr	Leu	Glu	Arg	Leu	Gly	Leu	Ser		
65		70						75						80		

ttc	cat	ttt	gag	acg	gag	atc	gaa	tac	aaa	atc	gaa	cta	atc	aac	gct	288
Phe	His	Phe	Glu	Thr	Glu	Ile	Glu	Tyr	Lys	Ile	Glu	Leu	Ile	Asn	Ala	
85		90						95								

gca	gaa	gac	gac	ggc	ttt	gat	ttg	ttc	gct	act	gct	ctt	cgt	ttc	cgt	336
Ala	Glu	Asp	Asp	Gly	Phe	Asp	Leu	Phe	Ala	Thr	Ala	Leu	Arg	Phe	Arg	
100		105							110							

ttg	ctc	aga	caa	cat	caa	cgc	cac	gtt	tct	tgt	gat	gtt	ttc	gac	aag	384
Leu	Leu	Arg	Gln	His	Gln	Arg	His	Val	Ser	Cys	Asp	Val	Phe	Asp	Lys	
115		120						125								

ttc	atc	gac	aaa	gat	ggc	aag	ttc	gaa	gaa	tcc	ctt	agc	aat	aat	gtt	432
Phe	Ile	Asp	Lys	Asp	Gly	Lys	Phe	Glu	Glu	Ser	Leu	Ser	Asn	Asn	Val	
130		135						140								

gaa	ggc	cta	tta	agc	ttg	tat	gaa	gca	gct	cat	gtt	ggg	ttt	cgc	gaa	480
Glu	Gly	Leu	Leu	Ser	Leu	Tyr	Glu	Ala	Ala	His	Val	Gly	Phe	Arg	Glu	
145		150						155						160		

gaa	aga	ata	tta	caa	gag	gct	gta	aat	ttt	acg	agg	cat	cac	ttg	gaa	528
Glu	Arg	Ile	Leu	Gln	Glu	Ala	Val	Asn	Phe	Thr	Arg	His	His	Leu	Glu	
165		170						175								

gga	gca	gag	tta	gat	cag	tct	cca	tta	ttg	att	aga	gag	aaa	gtg	aag	576
Gly	Ala	Glu	Leu	Asp	Gln	Ser	Pro	Leu	Leu	Ile	Arg	Glu	Lys	Val	Lys	
180		185						190								

cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca 624  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta 672  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat 720  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg 768  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

aaa tca aaa tta ccc tat gca aga gat cga gtc gtg gag gct tat gtt 816  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg 864  
 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat 912  
 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta 960  
 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa 1008  
 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat 1056  
 Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc 1104  
 Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg 1152  
 Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa 1200  
 Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa 1248  
 Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
 405 410 415

tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc 1296  
 Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
 420 425 430

gca aca tcg acc gct atg atc ggt cggt tat tgg aat gac acc agc tct 1344  
 Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392  
 Gln Leu Arg Glu Ser Lys Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt 1440  
 His Met Lys Glu Tyr Gly Leu Thr Lys Glu Ala Ala Ser Lys Phe  
 465 470 475 480

gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata 1488  
 Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc 1536  
 Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584  
 Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632  
 Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530 535 540

gtt gat gcc ata gtc ttt 1650  
 Val Asp Ala Ile Val Phe  
 545 550

&lt;210&gt; 15

&lt;211&gt; 550

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;400&gt; 15

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1                   5                   10                   15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20                   25                   30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35                   40                   45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50                   55                   60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65                   70                   75                   80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85                   90                   95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100                105                110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115                120                125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130                135                140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145                150                155                160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165                170                175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180                185                190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195                200                205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
210                215                220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
225                230                235                240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530 535 540

Val Asp Ala Ile Val Phe  
 545 550

<210> 16

<211> 1650

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: nucleic acid sequence encoding E-beta-farnesene synthase

<220>

<221> CDS

<222> (1)..(1650)

<223> Computer-generated nucleic acid sequence encoding peppermint E-beta-farnesene synthase protein

<400> 16

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Met	Ala	Thr	Asn	Gly	Val	Val	Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro	
1	5														15	

cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20	25													30		

ttt	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Leu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	
35	40													45		

gca	ctt	aag	caa	gaa	gca	aga	ggc	atg	ctt	atg	gct	gca	acc	act	cct	192
Ala	Leu	Lys	Gln	Glu	Ala	Arg	Gly	Met	Leu	Met	Ala	Ala	Thr	Thr	Pro	
50	55													60		

ctc	caa	caa	atg	aca	cta	atc	gac	act	ctc	gag	cgt	ttg	gga	ttg	tct	240
Leu	Gln	Gln	Met	Thr	Leu	Ile	Asp	Thr	Leu	Glu	Arg	Leu	Gly	Leu	Ser	

65	70	75	80	
				288
ttc cat ttt gag acg gag atc gaa tac aaa atc gaa cta atc aac gct Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala				85                    90                    95
				336
gca gaa gac gac ggc ttt gat ttg ttc gct act gct ctt cgt ttc cgt Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg				100                105                110
				384
ttg ctc aga caa cat caa cgc cac gtt tct tgt gat gtt ttc gac aag Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys				115                120                125
				432
ttc atc gac aaa gat ggc aag ttc gaa gaa tcc ctt agc aat aat gtt Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val				130                135                140
				480
gaa ggc cta tta agc ttg tat gaa gca gct cat gtt ggg ttt cgc gaa Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu				145                150                155                160
				528
gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu				165                170                175
				576
gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys				180                185                190
				624
cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala				195                200                205
				672
cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu				210                215                220
				720
ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Met Gln Asn Leu Tyr				225                230                235                240
				768
aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu				245                250                255
				816
aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val				

260	265	270	
tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg 864 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met			
275	280	285	
gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat 912 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr			
290	295	300	
gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta 960 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu			
305	310	315	320
gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa 1008 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys			
325	330	335	
atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat 1056 Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp			
340	345	350	
gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc 1104 Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr			
355	360	365	
gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg 1152 Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met			
370	375	380	
gaa agg cag cta ccg tca ttc caa gac tac gta aag aat acg gag aaa 1200 Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Thr Glu Lys			
385	390	395	400
acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa 1248 Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys			
405	410	415	
tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc 1296 Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu			
420	425	430	
gca aca tcg acc gct atg atc ggt cgg tat tgg aat gac acc agc tct 1344 Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser			
435	440	445	
cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392 Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe			

450	455	460	
cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe			1440
465	470	475	480
gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile			1488
485	490	495	
gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu			1536
500	505	510	
aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp			1584
515	520	525	
gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe			1632
530	535	540	
gtt gat gcc ata gtc ttt Val Asp Ala Ile Val Phe			1650
545	550		
<210> 17			
<211> 550			
<212> PRT			
<213> Artificial Sequence			
<400> 17			
Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro			
1	5	10	15
Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn			
20	25	30	
Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu			
35	40	45	
Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro			
50	55	60	
Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser			
65	70	75	80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
                   85                             90                         95  
  
 Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
                   100                         105                         110  
  
 Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
                   115                         120                         125  
  
 Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
                   130                         135                         140  
  
 Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
                   145                         150                         155                         160  
  
 Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
                   165                         170                         175  
  
 Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
                   180                         185                         190  
  
 Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
                   195                         200                         205  
  
 Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
                   210                         215                         220  
  
 Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
                   225                         230                         235                         240  
  
 Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
                   245                         250                         255  
  
 Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
                   260                         265                         270  
  
 Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
                   275                         280                         285  
  
 Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
                   290                         295                         300  
  
 Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
                   305                         310                         315                         320  
  
 Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
                   325                         330                         335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Thr Glu Lys  
385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

<210> 18  
<211> 1650  
<212> DNA  
<213> Artificial Sequence

&lt;220&gt;

<223> Description of Artificial Sequence: nucleic acid sequence encoding E-beta-farnesene synthase

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1)...(1650)

<223> Computer-generated nucleic acid sequence encoding peppermint E-beta-farnesene synthase protein

&lt;400&gt; 18

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Met		Ala	Thr	Asn	Gly	Val	Val	Ile	Ser	Cys	Leu	Arg	Glu	Val	Arg	Pro
1		5						10							15	

cct	atg	acg	aag	cat	gcg	cca	agc	atg	tgg	act	gat	acc	ttt	tct	aac	96
Pro	Met	Thr	Lys	His	Ala	Pro	Ser	Met	Trp	Thr	Asp	Thr	Phe	Ser	Asn	
20		25							30							

ttt	tct	ctt	gac	gat	aag	gaa	caa	caa	aag	tgc	tca	gaa	acc	atc	gaa	144
Phe	Ser	Leu	Asp	Asp	Lys	Glu	Gln	Gln	Lys	Cys	Ser	Glu	Thr	Ile	Glu	
35		40				45										

gca	ctt	aag	caa	gaa	gca	aga	ggc	atg	ctt	atg	gct	gca	acc	act	cct	192
Ala	Leu	Lys	Gln	Glu	Ala	Arg	Gly	Met	Leu	Met	Ala	Ala	Thr	Thr	Pro	
50		55			60											

ctc	caa	caa	atg	aca	cta	atc	gac	act	ctc	gag	cgt	ttg	gga	ttg	tct	240
Leu	Gln	Gln	Met	Thr	Leu	Ile	Asp	Thr	Leu	Glu	Arg	Leu	Gly	Leu	Ser	
65		70			75				80							

ttc	cat	ttt	gag	acg	gag	atc	gaa	tac	aaa	atc	gaa	cta	atc	aac	gct	288
Phe	His	Phe	Glu	Thr	Glu	Ile	Tyr	Lys	Ile	Glu	Leu	Ile	Asn	Ala		
85		90			95											

gca	gaa	gac	gac	ggc	ttt	gat	ttg	ttc	gct	act	gct	ctt	cgt	ttc	cgt	336
Ala	Glu	Asp	Asp	Gly	Phe	Asp	Leu	Phe	Ala	Thr	Ala	Leu	Arg	Phe	Arg	
100		105			110											

ttg	ctc	aga	caa	cat	caa	cgc	cac	gtt	tct	tgt	gat	ttt	ttc	gac	aag	384
Leu	Leu	Arg	Gln	His	Gln	Arg	His	Val	Ser	Cys	Asp	Val	Phe	Asp	Lys	
115		120			125											

ttc	atc	gac	aaa	gat	ggc	aag	ttc	gaa	gaa	tcc	ctt	agc	aat	aat	gtt	432
Phe	Ile	Asp	Lys	Asp	Gly	Lys	Phe	Glu	Glu	Ser	Leu	Ser	Asn	Asn	Val	
130		135			140											

gaa	ggc	cta	tta	agc	ttg	tat	gaa	gca	gct	cat	gtt	ggg	ttt	cgc	gaa	480
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

Glu	Gly	Leu	Leu	Ser	Leu	Tyr	Glu	Ala	Ala	His	Val	Gly	Phe	Arg	Glu	
145							150			155			160			
gaa aga ata tta caa gag gct gta aat ttt acg agg cat cac ttg gaa															528	
Glu	Arg	Ile	Leu	Gln	Glu	Ala	Val	Asn	Phe	Thr	Arg	His	His	Leu	Glu	
	165							170			175					
gga gca gag tta gat cag tct cca tta ttg att aga gag aaa gtg aag															576	
Gly	Ala	Glu	Leu	Asp	Gln	Ser	Pro	Leu	Leu	Ile	Arg	Glu	Lys	Val	Lys	
	180							185			190					
cga gct ttg gag cac cct ctt cat agg gat ttc ccc att gtc tat gca															624	
Arg	Ala	Leu	Glu	His	Pro	Leu	His	Arg	Asp	Phe	Pro	Ile	Val	Tyr	Ala	
	195							200			205					
cgc ctt ttc atc tcc att tac gaa aag gat gac tct aga gat gaa tta															672	
Arg	Leu	Phe	Ile	Ser	Ile	Tyr	Glu	Lys	Asp	Asp	Ser	Arg	Asp	Glu	Leu	
	210						215			220						
ctt ctc aag cta tcc aaa gtc aac ttc aaa ttc atg cag aat ttg tat															720	
Leu	Leu	Lys	Leu	Ser	Lys	Val	Asn	Phe	Lys	Phe	Met	Gln	Asn	Leu	Tyr	
	225					230			235			240				
aag gaa gag ctc tcc caa ctc tcc agg tgg tgg aac aca tgg aat ctg															768	
Lys	Glu	Leu	Ser	Gln	Leu	Ser	Arg	Trp	Trp	Asn	Thr	Trp	Asn	Leu		
	245					250			255							
aaa tca aaa tta cca tat gca aga gat cga gtc gtg gag gct tat gtt															816	
Lys	Ser	Lys	Leu	Pro	Tyr	Ala	Arg	Asp	Arg	Val	Val	Glu	Ala	Tyr	Val	
	260					265			270							
tgg gga gta ggt tac cat tac gaa ccc caa tac tca tat gtt cga atg															864	
Trp	Gly	Val	Gly	Tyr	His	Tyr	Glu	Pro	Gln	Tyr	Ser	Tyr	Val	Arg	Met	
	275					280			285							
gga ctt gcc aaa ggc gta cta att tgt gga atc atg gac gat aca tat															912	
Gly	Leu	Ala	Lys	Gly	Val	Leu	Ile	Cys	Gly	Ile	Met	Asp	Asp	Thr	Tyr	
	290					295			300							
gat aat tat gct aca ctc aat gaa gct caa ctt ttt act caa gtc tta															960	
Asp	Asn	Tyr	Ala	Thr	Leu	Asn	Glu	Ala	Gln	Leu	Phe	Thr	Gln	Val	Leu	
	305					310			315			320				
gac aag tgg gat aga gat gaa gct gaa cga ctc cca gaa tac atg aaa															1008	
Asp	Lys	Trp	Asp	Arg	Asp	Glu	Ala	Glu	Arg	Leu	Pro	Glu	Tyr	Met	Lys	
	325					330			335							
atc gtt tat cga ttt att ttg agt ata tat gaa aat tat gaa cgt gat															1056	

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp			
340	345	350	
gca gcg aaa ctt gga aaa agc ttt gca gct cct tat ttt aag gaa acc 1104			
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr			
355	360	365	
gtg aaa caa ctg gca agg gca ttt aat gag gag cag aag tgg gtt atg 1152			
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met			
370	375	380	
gaa agg cag cta ccg tca ttc caa gac tac gta aag aat tca gag aaa 1200			
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys			
385	390	395	400
acc agc tgc att tat acc atg ttt gct tct atc atc cca ggc ttg aaa 1248			
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys			
405	410	415	
tct gtt acc caa gaa acc att gat tgg atc aag agt gaa ccc acg ctc 1296			
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu			
420	425	430	
gca aca tcg acc gct atg atc ggt tat tgg aat gac acc agc tct 1344			
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser			
435	440	445	
cag ctc cgt gaa agc aaa gga ggg gaa atg ctg act gcg ttg gat ttc 1392			
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe			
450	455	460	
cac atg aaa gaa tat ggt ctg acg aag gaa gag gcg gca tct aag ttt 1440			
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe			
465	470	475	480
gaa gga ttg gtt gag gaa aca tgg aag gat ata aac aag gaa ttc ata 1488			
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile			
485	490	495	
gcc aca act aat tat aat gtg ggt aga gaa att gcc atc aca ttc ctc 1536			
Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu			
500	505	510	
aac tac gct cgg ata tgt gaa gcc agt tac agc aaa act gac gga gac 1584			
Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp			
515	520	525	
gct tat tca gat cct aat gtt gcc aag gca aat gtc gtt gct ctc ttt 1632			

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

gtt gat gcc gtc ata ttt 1650  
Val Asp Ala Val Ile Phe  
545 550

<210> 19  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<400> 19  
Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys

180	185	190
Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala		
195	200	205
Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu		
210	215	220
Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr		
225	230	235
Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu		
245	250	255
Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val		
260	265	270
Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met		
275	280	285
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr		
290	295	300
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu		
305	310	315
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys		
325	330	335
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp		
340	345	350
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr		
355	360	365
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met		
370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		

435

440

445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Val Ile Phe  
545 550

<210> 20

<211> 550

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>

<221> VARIANT

<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 20

Met Ala Thr Asn Gly Val Leu Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
 405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
 420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Tyr  
 530 535 540

Val Asp Ala Ile Val Phe  
 545 550

<210> 21  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>  
<221> VARIANT  
<222> {1}..(550)  
<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 21  
Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Gly Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
 405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
 420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
 515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
 530 535 540

Val Asp Ala Ile Val Phe  
 545 550

<210> 22

<211> 550

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:

E-beta-farnesene synthase protein

<220>

<221> VARIANT

<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 22

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
 1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn

20	25	30
----	----	----

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35                   40                   45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50                   55                   60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65                   70                   75                   80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85                   90                   95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100                105                110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Glu Lys  
 115                120                125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130                135                140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145                150                155                160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165                170                175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180                185                190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195                200                205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210                215                220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225                230                235                240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245                250                255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260                265                270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met

275	280	285
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr		
290	295	300
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu		
305	310	315
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys		
325	330	335
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp		
340	345	350
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr		
355	360	365
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met		
370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		
435	440	445
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe		
450	455	460
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Ala Ala Ser Lys Phe		
465	470	475
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile		
485	490	495
Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu		
500	505	510
Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp		
515	520	525
Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe		

530                    535                    540

Val Asp Ala Ile Val Phe  
545                    550

<210> 23  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>  
<221> VARIANT  
<222> (1) .. (550)  
<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 23

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1                    5                        10                        15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20                    25                        30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35                    40                        45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50                    55                        60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65                    70                        75                        80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85                    90                        95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100                    105                        110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115                    120                        125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130                    135                        140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Gly His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

<210> 24

<211> 550

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>

<221> VARIANT

<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 24

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
 1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
 20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
 35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
 50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
 65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
 85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Ser Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
 355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
 370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
 385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
 405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
 420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
 435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
 450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
 465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
 485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
 500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

<210> 25

<211> 550

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>

<221> VARIANT

<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 25

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys

115	120	125
<b>Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val</b>		
130	135	140
<b>Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu</b>		
145	150	155
<b>160</b>		
<b>Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu</b>		
165	170	175
<b>Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys</b>		
180	185	190
<b>Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala</b>		
195	200	205
<b>Arg Leu Phe Ile Thr Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu</b>		
210	215	220
<b>Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr</b>		
225	230	235
<b>240</b>		
<b>Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu</b>		
245	250	255
<b>Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val</b>		
260	265	270
<b>Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met</b>		
275	280	285
<b>Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr</b>		
290	295	300
<b>Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu</b>		
305	310	315
<b>320</b>		
<b>Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys</b>		
325	330	335
<b>Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp</b>		
340	345	350
<b>Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr</b>		
355	360	365
<b>Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met</b>		

370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
395		
400		
Thr Ser Cys Ile Tyr Ser Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		
435	440	445
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe		
450	455	460
His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe		
465	470	475
480		
Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile		
485	490	495
Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu		
500	505	510
Asn Tyr Ala Arg Val Cys Glu Ala Ser Tyr Thr Lys Thr Asp Gly Asp		
515	520	525
Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe		
530	535	540
Val Asp Ala Ile Val Phe		
545	550	

<210> 26  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

<220>  
<221> VARIANT  
<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 26

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Ala Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
225 230 235 240

Lys Glu Asp Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Asp Arg Asp  
340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355 360 365

Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met  
370 375 380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385 390 395 400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405 410 415

Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420 425 430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435 440 445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450 455 460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465 470 475 480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

<210> 27

<211> 550

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:

E-beta-farnesene synthase protein variant

<220>

<221> VARIANT

<222> (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

<400> 27

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gln Glu Ala Arg Gly Met Leu Met Ala Ala Thr Ser Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
 100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
 115 120 125

Phe Ile Asp Lys Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
 130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
 145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
 165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
 180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
 195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu  
 210 215 220

Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr  
 225 230 235 240

Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu  
 245 250 255

Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val  
 260 265 270

Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met  
 275 280 285

Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr  
 290 295 300

Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu  
 305 310 315 320

Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys  
 325 330 335

Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp  
 340 345 350

Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr  
355                   360                   365

Val Lys Gln Leu Ala Arg Ala Phe Asn Asp Glu Gln Lys Trp Val Met  
370                   375                   380

Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys  
385                   390                   395                   400

Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys  
405                   410                   415

Ser Val Thr Gin Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu  
420                   425                   430

Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser  
435                   440                   445

Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe  
450                   455                   460

His Met Lys Glu Tyr Gly Leu Thr Lys Glu Glu Ala Ala Ser Lys Phe  
465                   470                   475                   480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485                   490                   495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500                   505                   510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515                   520                   525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Val Val Ala Leu Phe  
530                   535                   540

Val Asp Ala Ile Val Phe  
545                   550

<210> 28  
<211> 550  
<212> PRT  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence:  
E-beta-farnesene synthase protein

&lt;220&gt;

&lt;221&gt; VARIANT

&lt;222&gt; (1)..(550)

<223> Computer-generated E-beta-farnesene synthase  
protein variant

&lt;400&gt; 28

Met Ala Thr Asn Gly Val Val Ile Ser Cys Leu Arg Glu Val Arg Pro  
1 5 10 15

Pro Met Thr Lys His Ala Pro Ser Met Trp Thr Asp Thr Phe Ser Asn  
20 25 30

Phe Ser Leu Asp Asp Lys Glu Gln Gln Lys Cys Ser Glu Thr Ile Glu  
35 40 45

Ala Leu Lys Gin Glu Ala Arg Gly Met Leu Met Ala Ala Thr Thr Pro  
50 55 60

Leu Gln Gln Met Thr Leu Ile Asp Thr Leu Glu Arg Leu Gly Leu Ser  
65 70 75 80

Phe His Phe Glu Thr Glu Ile Glu Tyr Lys Ile Glu Leu Ile Asn Ala  
85 90 95

Ala Glu Asp Asp Gly Phe Asp Leu Phe Ala Thr Ala Leu Arg Phe Arg  
100 105 110

Leu Leu Arg Gln His Gln Arg His Val Ser Cys Asp Val Phe Asp Lys  
115 120 125

Phe Ile Asp Asp Gly Lys Phe Glu Glu Ser Leu Ser Asn Asn Val  
130 135 140

Glu Gly Leu Leu Ser Leu Tyr Glu Ala Ala His Val Gly Phe Arg Glu  
145 150 155 160

Glu Arg Ile Leu Gln Glu Ala Val Asn Phe Thr Arg His His Leu Glu  
165 170 175

Gly Ala Glu Leu Asp Gln Ser Pro Leu Leu Ile Arg Glu Lys Val Lys  
180 185 190

Arg Ala Leu Glu His Pro Leu His Arg Asp Phe Pro Ile Val Tyr Ala  
195 200 205

Arg Leu Phe Ile Ser Ile Tyr Glu Lys Asp Asp Ser Arg Asp Glu Leu

210	215	220
Leu Leu Lys Leu Ser Lys Val Asn Phe Lys Phe Met Gln Asn Leu Tyr		
225	230	235
Lys Glu Glu Leu Ser Gln Leu Ser Arg Trp Trp Asn Thr Trp Asn Leu		
245	250	255
Lys Ser Lys Leu Pro Tyr Ala Arg Asp Arg Val Val Glu Ala Tyr Val		
260	265	270
Trp Gly Val Gly Tyr His Tyr Glu Pro Gln Tyr Ser Tyr Val Arg Met		
275	280	285
Gly Leu Ala Lys Gly Val Leu Ile Cys Gly Ile Met Asp Asp Thr Tyr		
290	295	300
Asp Asn Tyr Ala Thr Leu Asn Glu Ala Gln Leu Phe Thr Gln Val Leu		
305	310	315
Asp Lys Trp Asp Arg Asp Glu Ala Glu Arg Leu Pro Glu Tyr Met Lys		
325	330	335
Ile Val Tyr Arg Phe Ile Leu Ser Ile Tyr Glu Asn Tyr Glu Arg Asp		
340	345	350
Ala Ala Lys Leu Gly Lys Ser Phe Ala Ala Pro Tyr Phe Lys Glu Thr		
355	360	365
Val Lys Gln Leu Ala Arg Ala Phe Asn Glu Glu Gln Lys Trp Val Met		
370	375	380
Glu Arg Gln Leu Pro Ser Phe Gln Asp Tyr Val Lys Asn Ser Glu Lys		
385	390	395
Thr Ser Cys Ile Tyr Thr Met Phe Ala Ser Ile Ile Pro Gly Leu Lys		
405	410	415
Ser Val Thr Gln Glu Thr Ile Asp Trp Ile Lys Ser Glu Pro Thr Leu		
420	425	430
Ala Thr Ser Thr Ala Met Ile Gly Arg Tyr Trp Asn Asp Thr Ser Ser		
435	440	445
Gln Leu Arg Glu Ser Lys Gly Gly Glu Met Leu Thr Ala Leu Asp Phe		
450	455	460
His Met Lys Glu Tyr Gly Leu Thr Lys Asp Glu Ala Ala Ser Lys Phe		

465

470

475

480

Glu Gly Leu Val Glu Glu Thr Trp Lys Asp Ile Asn Lys Glu Phe Ile  
485 490 495

Ala Thr Thr Asn Tyr Asn Val Gly Arg Glu Ile Ala Ile Thr Phe Leu  
500 505 510

Asn Tyr Ala Arg Ile Cys Glu Ala Ser Tyr Ser Lys Thr Asp Gly Asp  
515 520 525

Ala Tyr Ser Asp Pro Asn Val Ala Lys Ala Asn Ile Val Ala Leu Phe  
530 535 540

Val Asp Ala Ile Val Phe  
545 550

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US98/20885

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :C07H 21/04; C12N 1/20, 9/88, 15/63, 15/70  
 US CL :435/232, 252.3, 252.33, 320.1, 320.1; 536/23.2, 23.6

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/232, 252.3, 252.33, 320.1, 320.1; 536/23.2, 23.6

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, STN: Medline, Caplus, Lifesci, Biosis, Embase, and Wpids

Search terms: Farnesene synthase

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	CROCK et al. Isolation and bacterial expression of a sesquiterpene synthase cDNA clone from peppermint ( <i>mentha x piperita</i> , L.) that produces the aphid alarm pheromone (e)- $\beta$ -farnesene. Proc. Natl. Acad. Sci. USA. November 1997, Vol. 94, pages 12833-12838, see abstract.	1-6, 9-14, 16-21, and 23-28
P,Y		----- 30
Y	SALIN et al. Purification and characterization of trans- $\beta$ -farnesene synthase from maritime pine ( <i>Pinus pinaster</i> Ait.) needles. J. Plant Physiol. 1995, Vol. 146, pages 203-209, see abstract.	1-6, 9-14, 16,-21, 23-28 and 30

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents:	*T*	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

11 JANUARY 1999

Date of mailing of the international search report

29 JAN 1999

Name and mailing address of the ISA/US  
 Commissioner of Patents and Trademarks  
 Box PCT  
 Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

NASHAAT T. NASHED

Telephone No. (703) 308-0196

**INTERNATIONAL SEARCH REPORT**

International application No.

PCT/US98/20885

**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  Claims Nos.: 7, 8, 15, 22, 29, and 31 because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:  

Claims 7, 8, 15, 22, 29, and 31 are drawn to specific amino and nucleic acid sequences. Applicants have filed the amino and nucleic acid sequences on a defective disket, and therefore, the data could not be entered into the data base to be searched.
3.  Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

**Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

**Remark on Protest**

The additional search fees were accompanied by the applicant's protest.



No protest accompanied the payment of additional search fees.